

ALZHEIMER'S DISEASE: THE ROLE OF EXERCISE AND MICROBIOME IN A TRANSGENIC MICE MODEL

PhD Thesis

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Abbreviations

AD	Alzheimer's disease
ANOVA	Analysis of variance
APP	Amyloid Precursor Protein
A β	β -amyloid
BBB	Blood brain barrier
CNS	Central nervous system
CV	Cardiovascular
CVD	Cardiovascular Disease
DSM	Diagnostic and Statistical Manual of Mental Disorders
ENS	Enteric Nervous System
GI	Gastrointestinal
GIT	Gastrointestinal tract
HIIT	High Intensity Interval Training
HM	Human Microbiome
HO- 1	Heme Oxygenase-1
HPA	Hypothalamic-pituitary-adrenal
HRP	Horseradish peroxidase
ISP	Ion Sphere Particles
LAB	Lactic acid bacteria
MCI	Mild Cognitive Impairment
MICT	Mild Intensity Continuous Training
MWM	Morris Water Maze
NEP	Neprilysin
NEP2	Neprilysin-2
NFT	Neurofibrillary tangles
NOR	Novel Object Recognition
NRF2	Nuclear factor erythroid 2-related factor 2
OFT	Open Field Test
OGG1	8 oxoguanine DNA glycosylase 1
oxoG	8-oxoguanine

PB	Phosphate buffer
PFA	Paraformaldehyde
PS1	Presenilin 1
PS2	Presenilin 2
ROS	Reactive oxygen species
SCFA	Short chain fatty acid
WT	Wild type

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1. Introduction

1.1. Alzheimer's disease

There is this old joke:

Grandpa: Grandson, do you know, what is the name of that German officer, who always hides my belongings?

Grandson: Yes, grandpa I know, that is Alzheimer.

Unfortunately, this is not a joke, Alzheimer's disease (AD) which was named after the German researcher –Alois Alzheimer–, who reported it in 1907, is the most common form of dementia. He described this condition as the patient was disoriented in time and space, she had serious breakdowns, memory impairments and she had difficulties with writing and speaking as well. Postmortem investigation has shown atrophic brain, with especially changed neuron tangles and a serious neuron loss in the upper layer of the cortex (1). Even though it is the cause of 60-80% of dementias, the underlying mechanism is one of the less understood ones, and regrettably, it is still without effective medical treatment (2).

It is not only the most frequent type of dementia, but it rises to one of the leading causes of death in western countries approaching the number of deaths caused by cardiovascular diseases (CVD) and cancer. Not the disease itself is so deadly, but it makes the body weak against illnesses. In most of the cases people with AD die from pulmonary diseases such as pneumonia and in this case, AD is not listed as the main cause of death. Because of this reason exact percent of death caused by AD itself is unclear. It is undoubtedly hard to precisely get AD diagnosed. In 2011 the National Institute on Aging has made new diagnosing criteria in which not only memory loss needs to be proved, but deeper data including biomarker tests are needed (3). Computed tomography scans can be performed to visualize beta-amyloid plaques but a simple and effective blood test to detect the disease with 100% certainty is still needed to be developed.

1.1.1. Clinical features

AD is a degenerative central nervous system disease which affects mostly the elderly population, namely people aged 65 or over. It is more and more commonly diagnosed as life expectancy increases, however, there is a tendency that in western countries the number of patients will be lower due to better cardiovascular (CV) prevention. There is an emerging evidence that poor heart function and CV state which is paired with lower cerebral blood circulation velocities and decreased ejection fraction can be a serious risk factor for developing AD (4). As the population ages, AD will put a higher burden to the society, not only medical wise but economically as well. It is estimated that by 2050 the number of patients with this disease will triple (www.alz.org). The fact that there isn't any medical treatment available which cures the disease, nor any that slows it down, elderly populations will be affected with AD in higher numbers. As a consequence of the disease, individuals will lose the capacity to live their life alone, they will need help continuously, twenty-four hours a day, seven days a week.

AD research has become popular in the last 30 years because governments realized that due to ameliorating medical treatments, the life expectancy increased, so did the number of patients with AD. Hence elevated financial inputs will be required to fulfill AD related care giving costs. Nowadays health care institutes are not prepared to supply this emerging need of patient care. Dementia is described as an acquired condition which has a serious impact on everyday activities and it consists of progressive cognitive impairment (5). Affected persons are not allowed to be left alone, because they are unable to complete everyday tasks such as dressing up, meal preparation or going on a short walk. AD shows different signs, firstly short term-, later long-term memory loss, struggle in day by day activities, speaking, writing and spelling difficulties, decline in problem-solving and navigation skills(6). As neuron loss and cerebral cortex shrinkage develops the ventricles become enlarged. Mostly right and left lateral ventricles are affected from this modification (7).

As time elapses elderly people need more personal assistance. It is usually realized by family members, but in some cases, it is not enough. A more reliable social

network is needed to fulfill the care giving needs. According to the latest Diagnostic and Statistical Manual of Mental Disorders (DSM) criteria dementia is a major neurocognitive disease (8).

AD has six stages, each stage usually lasts 5 years. It ranges from early symptoms until serious cognitive impairment and this process normally involves 20-25 years. When the disease begins, it is usually not diagnosed as AD, because symptoms can be the consequences for several other diseases and typical dementia-related signs are missing. Although this period could be the best for medical intervention, any product or method which could prevent Abeta formation, or limit the aggregation of β -amyloid ($A\beta$) could be disease modifying. As the symptoms are worsening, the disease will be more and more recognizable, but still it can be confused with other types of dementia. The fourth stage is the first one where mild cognitive impairment (MCI) can be investigated, and hippocampal atrophy is evident on a magnetic resonance imaging scan. Only with physical and pathological signs can be approved that it is AD, clinical diagnosis regularly happens in stage V (9). Usually when a patient goes to the doctor with the symptoms it is already too late, the disease is already in a progressive state. In the last phase cognitive decline arises from mild to severe, full-time surveillance is necessary. Typical symptoms are difficulties in eating and swallowing, inability to walk and to move, and the state of immune system becomes fragile.

1.1.2. Epidemiology

AD is the main cause of dementia, it accounts for 50-75% of all cases, affecting aging populations. (Figure 1.) In 2018, around 50 million people lived with dementia worldwide. The incidence is rising in low or middle-income countries while it shows a decreasing tendency in high-income countries. Still every 3 seconds a new case will be registered globally (10).

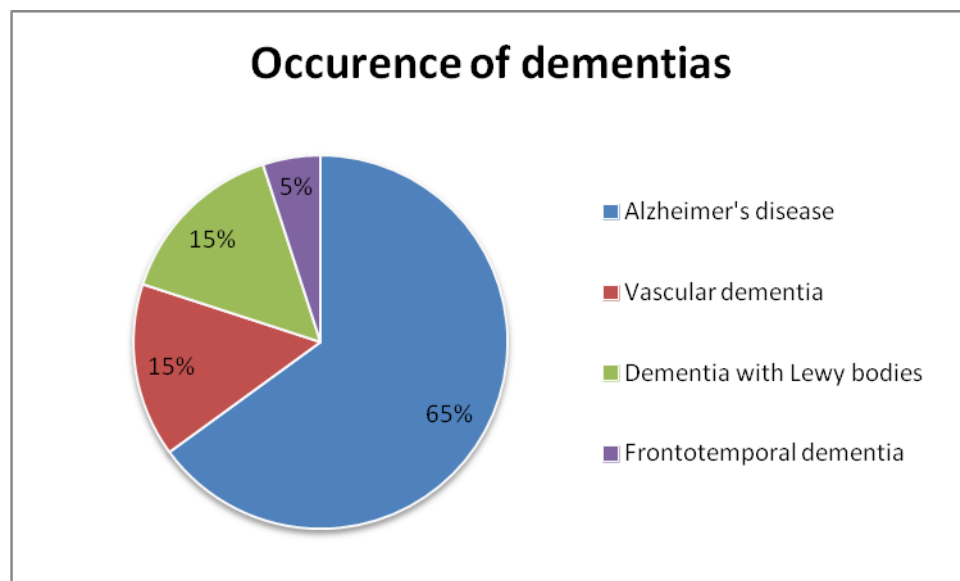


Figure 1. Percentage distribution of dementias

Occurrence of different types of dementia, showing that AD is responsible for the majority of the cases.

AD has two incidences. Sporadic or late-onset form is underlying 99% of the cases, it affects elderly people and it is due to a spontaneous gene mutation. While familial form is less common, it is hereditary, pathogenic mutations are mainly in genes APP, PS1, PS2 and it causes early onset dementia. AD is observed to be more prevalent in women than in man (11).

The reason for this gender dependent distribution is still unclear. Longer life expectancy for woman plays a role in it, or maybe because men have a higher mortality rate from other diseases. In the aging brain sex hormones have a crucial, defending role. Hormonal changes in females can also play a role in AD development. Estrogen

adjustment at the time of menopause may reduce the risk for AD. As for females the estrogen loss, the testosterone loss in males is also a normal, age-related phenomenon which automatically brings a higher risk for the disease (12).

1.1.3. Pathology- What are the molecules that are underlying AD?

It is still unclear why the disease develops, whether it is the intracellular amyloid deposition or the extracellular tau aggregates that cause AD or they are only consequences? The individuals suffering from AD have thinner cerebral cortex in the temporal, orbitofrontal and parietal brain regions compared to healthy individuals (13). Patients with this disease have senile plaques extracellularly mainly in the cortex and in the hippocampus, and neurofibrillary tangles (NFT) can be observed intracellularly. The amyloid cascade hypothesis (Figure 2. (14)) was originally declared back in 1992, suggesting that amyloid plaque deposition is the very first step to develop AD. Amyloid plaque depositions were declared as the main cause for the development of NFTs, cell loss and dementia (15). This hypothesis is partly accepted as an explanation for the developing disease. Familial AD evolves because abnormal formation and number of A β is present in the brain of the patients and combined with this, insufficient clearance takes place in disease specific brain regions. This imbalance can also be one of the factors underlying this disease. In contrast to this, it was proved, that in sporadic AD no correlation can be found between the plaque density and memory loss, still A β can be the initial step towards AD (16). So far scientific studies were unable to completely explore the mechanism underlying this disease.

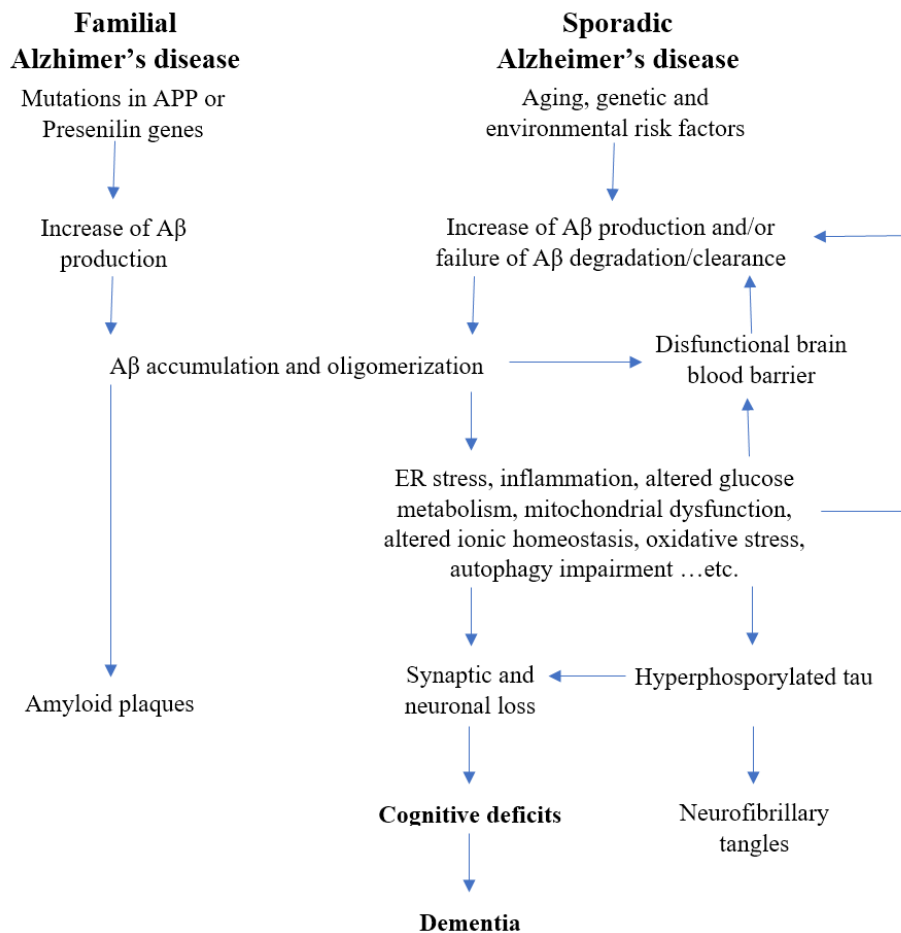


Figure 2. Amyloid cascade hypothesis

According to the amyloid cascade hypothesis Aβ accumulation and aggregation generates amyloid plaques and initiates a cascade mechanism (14). Later on this causes the dysfunction of BBB leading to inflammation, NFT formation and neuron loss. These changes are ending in cognitive impairment and dementia.

1.1.3.1. APP

The above mentioned hypothesis suggests that Aβ plaque deposition is crucial to develop the disease. Amyloid precursor protein (APP) is encoded on chromosome 21, and it is also present in healthy humans. It's neuronal function is still unknown, but it might have a role in synaptic plasticity (17). Aβ is the product of 2 cleavages made on APP, the first cleavage happens on the extracellular domain by β- secretase, the second which is made by γ-secretase happens on the transmembrane region (18). These 2

cleavages are necessary to form the 37-43 amino acid length A β (19). (Figure 3.) When APP has no mutation, it is usually cleaved by α -secretase resulting in a secreted APP α which is the amino terminal fragment of APP. Carboxy-terminal fragment is called CTF83 which is cleaved by γ -secretase but at the end, the remaining molecules P3 extracellularly and the amino-terminal APP intracellular domain is nontoxic (20). For example, Swedish mutation in APP initiates the cleavage by the β -secretase instead of α , resulting in A β that can aggregate in the extracellular space. It is suggested that extracellular deposits of A β are the triggering factor for neuroinflammation, microglial activation and also for tau tangle formation. Plaques are insoluble, and after their formation is finished, they are not so toxic. Latest inspections propose that soluble A β 40 and A β 42 which can be found in the plasma and central nervous system (CNS) of the patients trigger toxicity. Most mutations that affect APP are resulting in an elevated A β 42 to A β 40 ratio in the plasma, and the patient usually has an elevated presence of A β 42 in CNS (21).

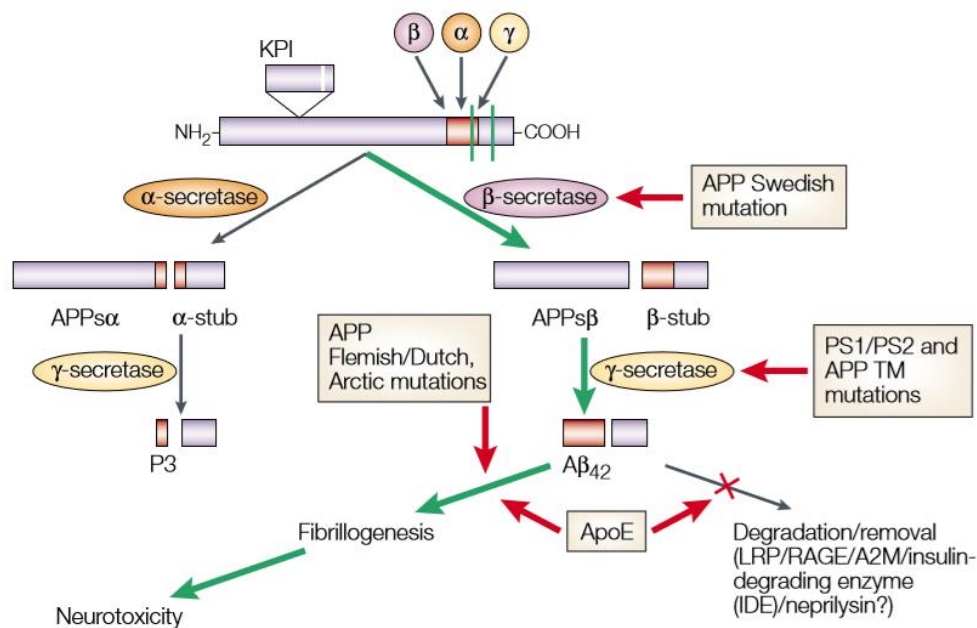


Figure 3. Route of amyloid processing

The route of amyloid processing (22) APP could be cleaved by α , β and γ secretase. In a healthy state cleavage happens initially by α - secretase followed by a cleavage made by γ secretase generating a final nontoxic 2 products, P3 extracellularly and a nontoxic APP fragment intracellularly. If Swedish mutation alters APP cleavage happens firstly by β -secretase followed by a γ secretase cleavage thus generating the toxic $A\beta_{42}$ which is able to aggregate and this mechanism will result in neurotoxicity.

1.1.3.2. Presenilin 1 and Presenilin 2

Presenilin 1(PS1) protein is the proteolytic subunit of the γ -secretase. Mice that overexpress a mutant PS1 are developing increased levels of $A\beta_{42}$. More than 50 PS1 gene mutations were discovered in patients with AD. Mutations in this gene are the biggest cause of early onset dementia. Presenilin 2 (PS2) also plays a role in APP cleaving, and also can be blamed for early-onset AD, still, it is only responsible for around 5 % of the cases while PS1 is responsible almost for 70% of the cases (23).

Mutations in PS1 and PS2 are resulting in an increased ratio of A β 42 to A β 40 as well as mutant APP.

1.1.3.3. Neprilysin and neprilysin-2

Insufficient A β clearance can be one of main causes of AD. Several proteolytic enzymes are found to take part in A β degradation, from which neprilysin is considered to be the most important one. Neprilysin (NEP) and neprilysin-2 (NEP2) are both zinc metalloendopeptidases and are part of the metalloprotease13 family (24). NEP is present in the periphery and in the central nervous system with a function of small peptide degrading (25). Even though NEP is responsible for A β degradation, elevated levels of this enzyme are still insufficient for A β elimination. Other NEP like enzymes such as NEP2 is also an important participant in A β degradation. It was shown that NEP2 knockout mice had elevated accumulation of A β species in the hippocampus (26).

1.1.3.4. ApoE

ApoE gene is the most important risk factor for familial AD. It has 3 alleles ApoE2 which is considered to be protective, ApoE3 is considered to be neutral and last but not least ApoE4 is considered to be a genetic risk factor for AD. In vivo and in vitro studies suggest that ApoE4 has an impact on A β clearance. (27). ApoE4 binds to A β peptides at residues 12-28, which can modulate A β accumulation, if this binding site is blocked amyloid plaque deposition can be prevented in the affected brain areas (28).

1.1.3.5. Tau protein

Tau protein is present in the healthy brain mainly in neurons, with the role of microtubule stabilization and polymerization. Abnormal phosphorylation of microtubule-associated protein tau results in a paired helical filament structured tau and NFTs. (Figure 4.) It was observed that hyperphosphorylated tau is present in the brain of patients suffering from AD, while healthy individuals do not have this mutation (29).

NFTs are one of the main pathophysiological characteristics of AD, they are responsible for the neuron loss. Phosphorylated tau appears in neurons well before tangle formation is present. This suggest that previous to disease recognition signaling pathways are already damaged (30). Mutations which affect tau function and/or isoform expression are more likely to get phosphorylated. Phosphorylated tau destructs microtubules and forms tangles, this modified structure can intrude axonal transport thus causing cell death (31). It was revealed that not only phosphorylation can convert tau into NFTs but truncation of the N or C terminal can also end in tau aggregation. It is still a question weather phosphorylated tau or truncated tau is more toxic to the cell (32).

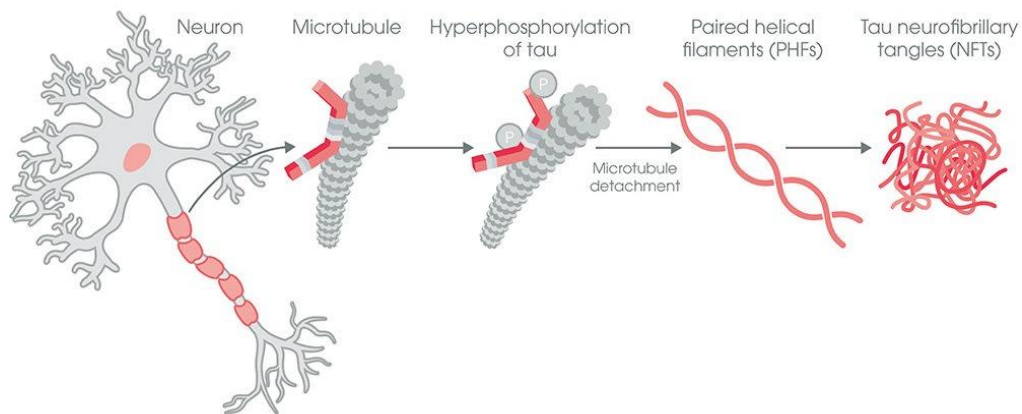


Figure 4. Neurofibrillary tangle formation

Tau protein takes part in microtubule stabilization. When it gets hyperphosphorylated, tau proteins destruct microtubules and aggregates forming NFTs (33).

1.2. Mouse models in AD

Mouse models can be used to study the pathology and progression of AD because they are able to mimic the effects of the disease such as A β plaques, tau tangles simultaneously with cognitive decline. Adversely none of these mouse models can represent other significant changes caused by the disease (34).

The first successful transgenic mouse model, using human APP was able to represent amyloid plaques within the mouse brain. Moreover, they have displayed several pathological features of AD. Model animals displayed a 5-14 fold higher A β presence than non transgenic littermate controls (35). Shortly after the first model, another human APP mutant mice was developed (36). After that presenilin mutant mice came to sight, to show that mutations in APP cleaving enzymes can also lead to A β accumulation (37). As knowledge expanded about AD the number of models representing different features of the disease increased. From the first transgenic model, scientific community arrived to a point where several different model types are available to choose from in close accordance with the investigation needs. Mice which can develop not only Abeta aggregates but tau tangles as well are available, allowing a more profound discovery on the mechanisms regulating the evolvement of AD.

Different model types are available according to the method of how the transgenic animal was made. Two different options can be used to gain a transgenic animal. The first option is to introduce a genetic mutation on top of existing genetic background. Second is to modify an existing gene of interest in its original position (38). Most common transgenic models contain the following modified genes: human APP, presenilin, tau and human ApoE (39). Even though numerous models and possibilities are available to study this disease in mice, unfortunately, these models cannot mimic exactly the disease pathology, as it happens in the human body. The main cause for this is that the immune system reacts differently in mice, it cannot recognize perfectly the human peptides applied in the models.

1.3. Oxidative stress in AD

Damage made by oxidative stress happens before the first hallmark of AD is present in the human brain (40). Neurons are more affected from oxidative stress than other cells in the brain because their metabolic rate is higher. Neurons on one hand contain polyunsaturated fatty acids, which enables them to be more responsive to reactive oxygen species (ROS), generating lipid peroxidation and molecular destruction. On the other hand glutathione content, which is a known antioxidant is really low in neurons (41). Oxidative stress contributes to lipid, protein and/or nucleic acid oxidation, mitochondrial dysfunction, metal accumulation, tau pathology and inflammation (42). Mitochondrial dysfunction is undoubtedly one of the main ROS producer in AD, with signs of abnormal mitochondrial axonal trafficking, and with disrupted glucose metabolism. These problems lead to an elevated ROS production which contributes to early stages of AD. However there are several factors which contributes to ROS production, there is no available antioxidant cure to prevent oxidative damage in AD (43).

Nuclear factor erythroid 2-related factor 2 (Nrf2) is an important gene taking role in the fight against oxidative stress. Nrf2 levels are usually low in AD. It was observed that Nrf2 regulates other enzymes that are taking part in redox regulation such as heme oxygenase-1 (HO-1) and glutathione cysteine ligase modulatory subunit. Elevated expression of Nrf2 in AD models reduces oxidative stress moreover if it was overexpressed in the territory of hippocampus it results in better memory and learning. On top of that the activation of HO-1 improves learning and memory in mouse models of AD (44).

DNA damage caused by oxidative stress is also a main symptom in AD. Oxidative DNA damage removal is accomplished by 8-oxoguanine DNA glycosylase 1 (OGG1) (45). The level of this glycosylase is significantly lower in AD patients than is controls (46). OGG1 is involved in the first step of base excision repair, which is the most common pathway in DNA damage removal. It excises 8-oxo-2'-deoxyguanosine from DNA. OGG1 enzyme has numerous variants and it seems that there is a great significance of which genotype is present in the plasma of AD patients (47).

1.4. Microglial activation in AD

Microglia is a phagocyte within the CNS and plays a critical role in A β degradation, it can interact with the soluble forms and also with the insoluble forms of A β . This interaction is different in both cases (48). Microglia is found to be in close relation with amyloid plaques showing an activated phenotype. Their size and number is in correlation with plaque size. Even though they are phagocytes, CNS resident microglia is unable to remove A β plaques with phagocytosis (49). Uncontrolled microglia activation can lead to chronic inflammation and to the release of several inflammatory cytokines thus causing neurodegenerative diseases such as AD. Still these cells are the only ones to mediate immune responses in the CNS. In addition to inflammatory cytokine production, microglia is able to produce anti inflammatory cytokines and reactive oxygen intermediates (50). In late onset AD there is emerging evidence that activated microglia is responsible not only for continuous inflammation but also for neurodegeneration and synapse loss. In AD microglial activity has two faces. Firstly they are cells responsible for brain health and tissue homeostasis with a sufficient A β clearance serving as a protective factor against AD. Secondly if A β accumulation occurs and microglial activation is continuous, neuron loss and inflammatory cytokine production happens helping the disease to develop. These results suggest that microglial activation as a therapeutic strategy is only suitable in the early stages of AD (51).

1.5. Methods of prevention

So, what can be done to prevent this disease? Not much in fact. It is believed that regular physical exercise, a healthy diet, and continuous learning can help. Several types of research have proved that physical exercise as for the body and as for the brain can be beneficial. Lifelong learning can keep the mind refreshed, and in good condition. Also, it is believed that a healthy diet can help to postpone the disease onset. It is observed that people who received better education and live in better circumstances are less affected by AD.

1.5.1. Role of exercise

The effects of regular exercise have been in focus of research for a long time, and it is proved to be beneficial in several diseases and in numerous health conditions. Practiced at any age, for any duration and in any type, it is hard to say any disadvantage about exercising. It can help fight against negative stress, create an esthetic body, establish a more positive thinking, keep a healthy lifestyle and on top of all that it can prevent numerous diseases. There are some obvious examples such as high blood pressure, diabetes and excess weight caused health problems like back and knee pain, insomnia and hampered respiration where exercise can be a reassuring solution. Exercise can also cause some not so obvious changes in the body, it is liable for neurogenesis in the hippocampus which is the most important area in the brain for learning and memory forming (52). Also, it can ameliorate the survival of newborn neurons in the dentate gyrus (53).

Activity of OGG1 is also upregulated by the effects of exercise thus reducing DNA damage (45). This enzyme is responsible for the repair of 8-oxoguanine (8-oxoG), the most abundant DNA lesion caused by oxidative stress. These lesions become more abundant with age, and they can be responsible for inflammation, and can be related to CNS diseases such as AD, and Parkinson's disease (54). Interestingly, exercise is not altering the ROS levels in the brain (55).

It is believed that regular physical activity can also ameliorate CNS related diseases, not just memory improvement but it could delay the progression of AD as well as lower the incidence of it (56). Furthermore, exercise can reduce the risk of developing AD (57). However the exact mechanism underlying this phenomenon is still unclear, it is believed that exercise can increase the metabolic function of the brain and can reduce the effect of oxidative stress in AD (58).

There are several studies that prove that voluntary or forced exercise can slow down disease progression and can amend physical symptoms caused by the disease. Voluntary exercise is known to have more beneficial effects than forced exercise however it is mainly because of stress-related factors. Voluntary and forced exercise differs in training time and in running speed. If the goal is, to achieve the same distance during the training period, animals which were allowed to run voluntarily, accomplished

the distance with several short running sessions for a faster speed, than animals that were forced to run. At the end, voluntarily exercising animals reached the distance goal first. At the end of training months, only anxiety related experiments showed an advantage for animals in the voluntary exercise group. In open field test these animals were more active and crossed more inner areas, while there weren't any differences neither in Morris water maze test (MWM) nor in body weight measurements (59).

Interval training is a time efficient way to do daily exercise. It is believed that high-intensity interval training (HIIT) is a more efficient way for weight loss than moderate intensity continuous training (MICT). HIIT is composed of two different intensity sections, one of them is a high-intensity period, and the other is low-intensity period. These two sections are changing during the training period creating more challenging cardiovascular training. It turned out that this method is more efficient only if we take time into consideration (60). It has no more beneficial effects than MICT in weight loss or in body composition.

Treadmill running reduced A β 42 levels in the brain of Neuron specific enolase/APPswe tg mice, also it reduced the escape latency time in the MWM compared to the sedentary group (61). As a prevention method in APP/PS1 mice, regular voluntary physical activity is proved to slow down the disease development (62). It has also been observed, that forced treadmill running can significantly improve the memory deficit and the learning abilities of APP/PS1 transgenic mice (63). When exercise was used in a progressed, irreversible state of AD, unfortunately no improvement was shown in the cognitive function of the animals due to the training. There was no improvement in the escape latency time in MWM nor decreased number in the A β plaque burden in motor-related brain regions of APP/PS1 mice at the age of 10 months (64). It seems that in the progression of AD exercise alone cannot play a protective role, maybe human microbiome can have a similar retentive effect on disease pathology. It is well known that exercise can reversibly alter the microbiome, not only the composition but the function as well (65). The only question is whether changes in microbiome can help delay the onset of the disease?

1.5.2. Effects of environmental enrichment

The role of environmental enrichment is controversial, because this type of housing differs from conventional housing. Cage with enrichment could contain a running wheel, toys for the animals and sometimes the cage has a bigger area collectively. Thus, it is hard to observe the effects of environmental enrichment solely. Positive effects regarding enrichment consistently come from the beneficial effects of exercise. In mouse models of AD it was proved that enriched housing results in reduced A β level in the brain together with decreased number of senile plaques. Moreover, the enzyme responsible for A β degradation, neprilysin, was observed in a higher quantity (66). Both factors together, physical activity and environmental enrichment can trigger cognitive improvement in models of CNS diseases (67). Even so environmental enrichment alone may induce synaptogenesis, and can have beneficial effects on synaptic plasticity (68).

1.6. Microbiome

The human microbiome (HM) was not in the center of research until very lately. In the last decade there is emerging evidence that the microbiome is far more essential than it was imagined earlier. Its composition and its function can be the underlying cause for numerous illnesses. HM consists of various participants such as archaea, protozoa, viruses, eukaryotes, and mainly bacteria. These organisms inhabit different parts of our body for instance skin, mouth, vagina, oesophagus and gut where they are present in a different composition. The biggest part, around 95% of the HM is in our gastro intestinal (GI) tract which plays a major role in nutrition, inflammation, immunity and its composition can affect neuronal function(69). Although the content of the microbiome not only differs from person to person, from age and from the ethnical origin, the main characteristics can be generalized and observed. It was believed for a long time that microbes in our body are outnumbering the number of human cells in a tenfold magnitude.(70) A recent study from 2016 revealed that this ratio is almost 1:1, to be exact is 1.3:1 (71). If we consider gene number this ratio is even much higher

150:1 (72) which is such an enormous quantity, that it makes complicated to investigate every participant and every function of the HM. Lately gut microbiome research became a hot topic, numerous researchers suggest that there must be a lot on our gut microbiome if we talk about health or disease. Functional relevance of the microbiome is really wide, it is fundamental for normal GIT (Gastrointestinal tract) motility, digestion and host metabolism, it takes part in the maintenance of barrier function, it is responsible for epithelial cell repair after injury, it takes part in immune function and pathogen recognition (73). A bidirectional pathway is suggested to exist between the gut and the brain in contrast to the previously believed only one direction pathway, that CNS regulates the enteric nervous system.

1.6.1 Microbial changes during life

The human gut microbiome is an interesting and continuously changing environment. During pregnancy, in the intrauterine life, a part of the maternal microbiota is already inherited through the placenta. *Lactobacillus* and *Bifidobacterium* DNA was isolated from human placenta suggesting an evidence against the uterus being sterile (74). When a baby is born, normally it inherits the microbiome from the mother's vaginal microbiome, which composition allows easier breast milk digestion. In contrast to this if someone was born through C section, the primary bacterial colonization happens from the mother's skin flora. In newborns the majority of bacteria which colonizes the GI tract is Lactobacilli, whose function is to prepare the environment in the gut for further bacteria until total GI maturation is reached (75). Later on, bacteria are obtained from the environment. It is observed that the microbiome of babies fed by breast milk is mainly different from the microbiome of babies fed by formula. If the formula contains prebiotics, the microbiome composition of formula fed babies can be more similar to the microbiome of babies fed by breast milk (76). Gut microbiome changes a lot within the first 3 years of age, the number and the variety of species become larger. The GI tract composition can mirror the food intake as well as antibiotics can transform the original composition of it. (Figure 5.) These differences can be observed even several years later since early life microbiome is the foundation of

the adult microbiome. It is suggested that aberrant microbiome between ages 0-3 years can lead to diseases, most probably due to the altered development of immune system (77). After early years the gut microbiome reaches a balanced state, it still has shifts and changes, but it is more stable, renews and adapts time to time during adulthood. Microbial shifts can occur due to an illness, antibiotic usage, or even due to new type of diet, but normally the HM can remain stable for months in some cases even for years (78). The composition of the GIT on its functional level doesn't change a lot during a lifespan. On the contrary on species level the actual composition transforms doubtlessly. Changes in the GI tract are slowing down by age 65, elderly people have less diverse microbiome and differences between individuals are blanching (79).

The gut microbiome is a self regulating system, the human body attempts to keep it in a balanced state, maintaining all necessary functions, without diverging from the original setup. Each person has a 150-400 species in their gut, which is mainly composed of 4 phyla: *Firmicutes*, *Bacteroidetes*, *Actinobacteria* and *Proteobacteria*. Individual bacterial diversity depends on age, diet, weight and geographical factors (80). If the microbiome of two individuals is compared, a huge difference can be observed between them. Interestingly, during a one year observation period, the composition of the microbiome of the individuals do not varies significantly so the differences between the 2 individuals will not disappear or become even more different. In contrast to the microbiota stability, it can be observed how external factors have a huge role in the microbiome formation. Traveling to a different country, consumption of food from different gastro culture and gut infections can easily destabilize the gut balance. These changes can perturb the structure up to the phylum level. Reversibility of perturbation can have 2 types, environmental and community disturbance model. When environmental perturbation happens, changes being done in the microbiome will go back to the original structure when the environment goes back to the original state, for example when we return home after a long journey. Community perturbation results in altered microbiome composition which can be stabilized, and later on, when perturbation vanishes this newly formed community remains stable and functional stability can become permanent (81).

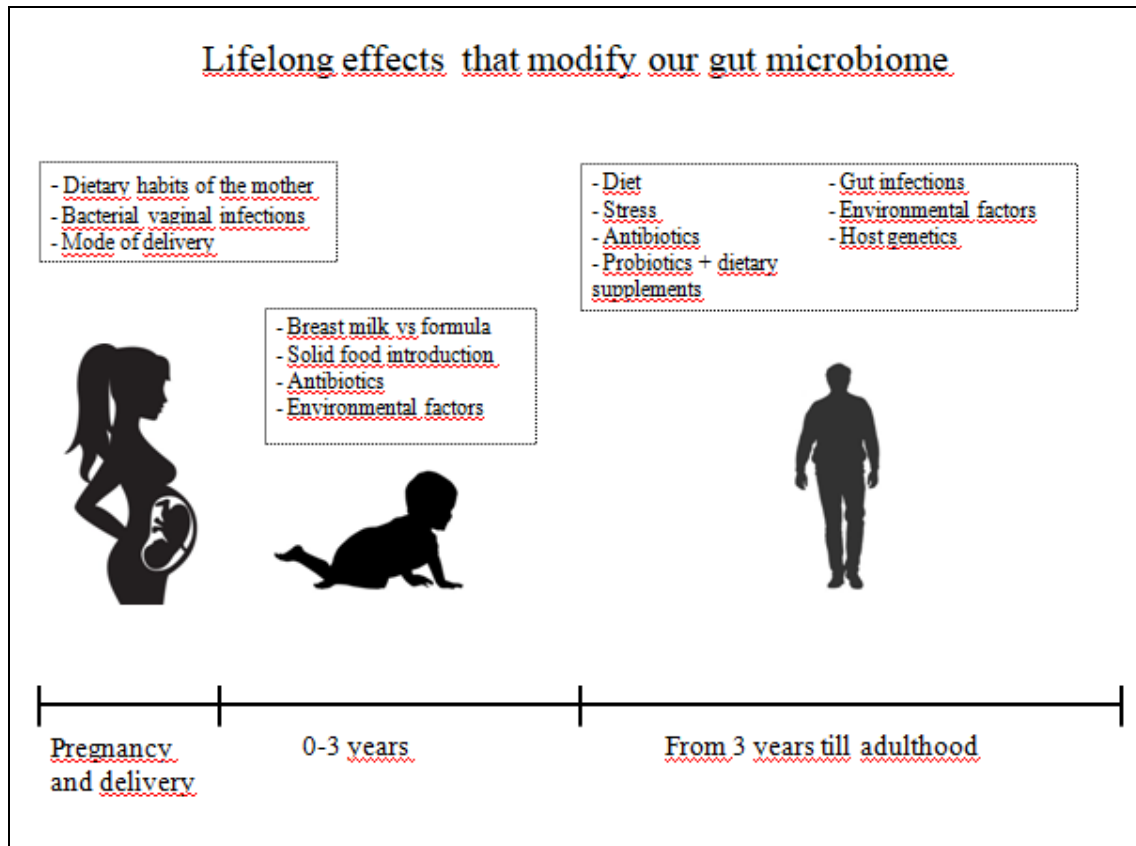


Figure 5. Lifelong effects that modify our microbiome

It is easily noticeable, that a set of factors are responsible for the gut microbiome development. Most aggressive effects are affecting the microbiome in the first 3 years of our life.

1.6.2. Healthy host means a healthy gut?

As it was mentioned previously, every individual has a continuously changing microbiome, but somehow this system is still able to keep a balanced state. Sometimes, this stability is broken. How can it be known, that for the proper microbiome function help is needed to restore the balanced state? Is it possible to know, what should be done to improve the state of the gut microbiome? What can be considered as a healthy microbiome? If the host is healthy does that mean that the host microbiome is healthy too, or the host can be healthy only if the gut is healthy? It was proven before that the

state of microbiome can affect the central nervous system of the host. It was observed on patients suffering from autism or Parkinson's disease (82) (83) (84).

If the microbiome is considered to be healthy, a higher and thicker mucus layer is observed together with higher short chain fatty acid (SCFA) and antimicrobial signal production. This elevated SCFA level helps to reduce food intake and improve glucose absorption (85). Main SCFAs produced by gut bacteria are butyrate, propionate and acetate from which butyrate is the best known for its beneficial effects. After a diet rich in fiber elevated levels of butyrate and acetate were observed together with reduced appetite and with improved glucose tolerance. While increased butyrate level is beneficial to the host, increased fecal propionate level predicts a higher risk for type 2 diabetes (86). On the contrary, low diversity of the gut microbiota can be in association with an unwell health state including allergy, obesity, bowel diseases and even neurological disorders. For a healthy body the gut microbiome must be in contact with the host's immune system (87). When the microbiome is investigated it is essential to measure not only the presence of several bacterial species but their activity as well. The activity measurement is not that easy as it seems. In one hand there is no easy and cost efficient way to quantify the metabolite production of all bacterial species. On the other hand the microbiome of every individual should be examined separately. There is no current understanding on how the activity of microbes vary within the same disease (88). Recent findings ensure that every person has a unique microbiome, it presents a new possibility to develop personalized medications. Individual medication usage will bring less side effects, reduced disease risk together with a more effective, non invasive and affordable disease treatment (89). Considering these facts, it is impossible to declare what kind of microbiome composition can be stated as a healthy microbiome.

1.6.3. Leaky gut

Leaky gut is a common and summarizing nomenclature for increased epithelial permeability in the GI tract, most commonly because of mucosal damage caused by stress or inflammatory factors. Stress factors can be endurance training, non-steroidal anti-inflammatory drug administration and pregnancy. These factors can cause mucosal

damage and modify intestinal permeability. Stress caused damage can be reversed by an appropriate diet without any pharmacological intervention. On the contrary damage caused by some inflammatory event such as allergy, ageing, CNS disorders cannot be restored with diet because the intestinal barrier function normalization will not repair the modification. Still it is not proven that increased gut permeability can cause any serious problems (90). Mucosal permeability may activate a mucosal immune response. This activation can depend on the severeness of mucosal deregulation and also on the circumstances. It seems that the state of mucosa is in active connection and cooperation with the immune system so inflammatory response is not obligatory (91). One of the most common bacteria in the gut is *Bacteroides thetaiotaomicron* which is a well known acetate producer, it hydrolyzes polysaccharides delivered from diet or from the host and it increases goblet cell differentiation. Under normal circumstances it is part of the normal gut flora and it can promote mucus production. If an acetate consumer and butyrate producer bacteria is present in association with *Bacteroides thetaiotaomicron* this effect can diminish and can lead to thinner mucus layer (92). Acetate overproduction in the GIT usually happens if a diet high in fat and calorie is followed. Acetate can trigger an elevated insulin secretion in the gut causing insulin resistance and later on obesity (93). Also it can promote the secretion of a hunger hormone called ghrelin which can cause increased food intake creating a continuous loop of eating, acetate overproduction, ghrelin production and then eating again (94). It is suggested that leaky gut can underlie some CNS disorders by influencing blood brain barrier function (BBB). If this is so, small bacterial components can pass the BBB (95).

1.6.4. Gut-brain axis

It was observed in the last century that emotional changes can have different effects on the GI tract. But it is a recent finding that alterations in microbiota composition can affect brain function. It is suggested that variations in the microbiome can also have several effects on distinct diseases such as autism, anxiety, depression, irritable bowel disease and even memory related dysfunctions can be due to the microbiome. Gut-brain axis is a bidirectional connection between the CNS and enteric

nervous system (ENS). Microbes not only have effects on ENS but on CNS as well. Communication is mediated through neural, neuroendocrine, and neuroimmune pathways. Neuroendocrine pathways are mediated through the hypothalamic-pituitary-adrenal (HPA) axis. For the normal function of this axis the development of an appropriate stress response is essential. This stress response is largely dependent on the normal gut colonization in early life (96). Since it is known that early microbial composition can have an effect on CNS development and function, there is urging need to find the mechanisms and patterns responsible for this phenomenon (97). Most of the findings are based on animal models, mainly with germ free mice where anxiety related behavior is reduced compared to conventional mice (98). Not just antianxiety- but antidepressant like behavior was observed in germ free mice (99). First results arrived when stress related factors were investigated, but there is huge hope to find relations between dysfunction of gut microbiome and gut disorders such as irritable bowel disease (100). Nervus vagus was the first hypothetical connection between these two interfaces, and it is partly true that it has an important role in the communication. It is still unclear if the bacterial composition of gut is important when we talk about the effects on CNS, or the secreted metabolites are the responsible for all the changes. The nervous system is able to detect an infection without any immune response just because a pathogen is present in the gut. Also some neuropsychiatric disorders can be improved with antibiotic registration. In these cases there is no proven role of the secreted metabolites, only the presence or absence of bacteria is important. When we talk about metabolites it is even a more complex topic. There is no comprehensive knowledge available about what type of metabolites are secreted from specific bacteria. One type of metabolite can be a product of several different bacterial species, and metabolite secretion can be altered by environmental factors such as diet or infections(101). If there will be a reliable method to investigate bacterial metabolite secretion, most probably that will be the route to follow in the scientific exploration of gut-brain axis.

1.6.5. Microbiome and exercise

If the role of exercise on the microbiome is investigated, several evidences can be found suggesting that physical activity can modify the composition of the microbiota. Various questions are arising such as what kind of exercise is able to initiate modifications, how long does it need to be made, what age is optimal to start exercising, how is possible to perceive the changes caused by activity? Will anything be observed outside like bigger muscles, or quicker performance development, or only inside differences will be made such as improved nutrition utilization?

If only training effects are taken into consideration, it is a challenging task to investigate its impact on the microbiome, still there is an arising need to specify the results achieved by workout. It has been reported that physical activity during childhood is able to make the microbiome more diverse and can promote the growth of butyrate producing bacteria. Furthermore, exercise in childhood has a higher influence on microbiome modification than in adulthood. Changes made in childhood can last until adulthood, while changes occurred in adulthood are considered to be more unstable. These changes might affect body fat content and skeletal muscle mass furthermore in the long run metabolic health can be improved (102). Even though exercise in childhood has enormous beneficial effects on health state, the existing positive effects of regular physical activity in the elderly population cannot be forgotten. It can raise bacterial diversity which may result in a better immunological state, together with an exercise driven anti-inflammatory effect (103). If the fecal samples of sedentary or control animals are compared, it can be observed that their microbiome composition is undoubtedly different, samples from the exercise group showed a higher n-butyrate concentration (104). N-butyrate is a well known SCFA which has an inhibitory effect on tumor development, also its low levels are observed in inflammatory bowel disease. One of its main roles is to modulate NF- κ B activation (105). Elevated butyrate levels were also observed in individuals with a better cardiovascular fitness state, if level of VO_2 max is higher the amount of produced butyrate is also higher (106). Butyrate is secreted by bacteria when dietary fiber is processed. Butyrate is essential in the colon, it produces about 70% of the energy used by colonocytes. It has anti-inflammatory effects

and it induces regulatory T-cell proliferation. Indeed, SCFAs are important mediators against inflammation. SCFAs are fatty acids that contain less than six carbon atoms, most common forms are butyrate, propionate and acetate. They take part in fiber digestion, and they are important molecules because they can cross the BBB. In a mouse model of AD it was presented that the number of SCFA in the fecal samples was significantly lower than in control animals, which can result in the perturbation of several metabolic pathways, therefore opening new possibilities to therapy (107). Most probably SCFAs can mediate the effects of probiotics and prebiotics, and they can be involved in the communication between the GIT and CNS (108).

1.6.6. Microbiome and diet

Diet is one of the main modulators of the microbiome. Dietary intake alters the microbiome on a daily basis. It can modify its components on both the short and long run. A one-off change can have a serious impact on its state, while a continuous life change can modify the roots of the microbiome composition. It is believed that human nutrition intake is mainly dependent on gut bacterial composition, namely the food preferences are in accordance with bacterial needs. Specific carbohydrates are proved to be the main nutrition to them. Some of the food cravings should be due to our microbiome needs, microbes may manipulate our eating behavior in favor to their advantage. It is suggested that due to the fight for nutrition and habitat in a higher population diversity may lead to a more colorful composition without the opportunity that any particular participant can overgrow the others, resulting in an easy host dietary manipulation (109). There is also an abundant number of evidence that microbiota of obese and lean individuals is different, if their microbiome is exchanged the obese becomes lean and vice-versa. Can the use of pro- and prebiotics modify the pattern of our microbiota? Probiotics are live microorganisms mostly from phyla *Lactobacillus* and *Bifidobacterium* often referred to as healthy bacteria because they provide health benefits. Prebiotics are substances that support the growth of probiotics (110). Some lactic acid bacteria (83) is able to produce water soluble vitamins mainly from the group of vitamin B. These are mainly folic acid, riboflavin and B₁₂ vitamin. Some members of

Lactobacillus genus can have a positive effect on the host health state (111). Riboflavin is essential for cellular metabolism, it serves as a coenzyme for hydrogen carrier molecules in redox reactions. Riboflavin is produced by *Bacillus subtilis* (112). The de novo synthesis of B₁₂ vitamin is not very common in the human intestine, only few bacterial strains are able to do that. One of them is *Lactobacillus reuteri* (113). B12 vitamin is essential for healthy neurological development, deficiency in this vitamin may lead to cognitive decline including AD, depression and stroke (114). These observations are suggesting a new methodology in medical sciences. What if every disease can be cured through the GIT? What if the GIT can be colonized with beneficial bacteria? Will the average health state become better? Shifts in microbial composition can affect behavior, can have an effect on mood and maybe it can make changes in the CNS.

1.6.7. Microbiome and AD

Microbiome most probably has a role in AD as well. One of the main products of bacteria is different amyloids which can underlay amyloid accumulation in the brain (115). It was investigated in numerous studies what effects can be observed if probiotics were administered to patients with AD. Since AD usually occurs at a senior age when diet is changed and it turns more one-sided, instability in microbiome composition occurs, thus leaky gut syndrome can develop which can easily lead to unexpected inflammation. Probiotic supplementation can activate the immune response to inflammation in patients with AD (116). Furthermore, it was observed that not only probiotic supplement composition matters but the duration of regular use of probiotics is also important. There is no evidence that probiotics can help in severe stages of AD. Meanwhile it was shown that in early stages this kind of supplementation can lead to an increase in some antioxidant factors, and a slight improvement in cognition. Further investigation is needed about the longitude of administration, and about the most effective composition of the supplement (117). Healthy microbiome is fundamental for the integrity of BBB (118). Stress can have harmful effects on the microbiota and thus on the brain, these modifications can cause a more severe impact on ageing brain (119).

Aging and stress together can weaken the gastrointestinal barrier as well. Unfortunately a detailed analysis of microbiome of patients with AD is still lacking, but several evidence suggest that the composition of microbiome can play a bigger role on the disease progression than it was believed previously (120).

Seven dietary and lifestyle guidelines were proposed to help prevent AD at the International Conference on Nutrition and the Brain, from which 6 relate to dietary habits and 1 relates to regular exercise (121). (Table 1.)

Table 1. Dietary and lifestyle guidelines to prevent AD.

<u>Possible dietary and lifestyle guidelines to prevent Alzheimer's disease</u>	
Reduce	Add
Minimize saturated fat intake	Instead of meat and dairy products vegetables, fruits and whole grains should be eaten
Reduce iron consumption	Dietary intake of B12 vitamin is necessary
Avoid the use of materials which contain aluminium	Do aerobic exercise at least 3 times a week
	Vitamin E should come from food instead of dietary supplements

2. Objectives of the study

The main objective of the study was to investigate if interval treadmill running and specific probiotic lysate supplementation can delay the onset of dementia in APP/PS1 transgenic mice.

We have found several publications with the positive effects of marathon type exercise on the development of AD and we were interested if high intensity interval exercise can have similar positive effects on the cognitive functions. Moreover it was proved that higher running speed results in greater metabolic demand for the brain. Because we wanted to test interval training effects, which relies on continuous speed control we could not use voluntary exercise model.

We also wanted to reveal that a specific probiotic lysate supplementation can significantly modify the composition of the gut microbiome. And if yes, can these changes make such a big difference in cognition? Is it possible that changes in microbiome can enlarge the positive effects of exercise?

It was at the center of our interest that these two types of treatments can be effective alone, or they can have more beneficial effects if they are applied together. Is it possible, that these preventive methods can postpone the development of AD?

Hypotheses:

1. Regular physical activity and probiotic lysate treatment will enhance the cognitive function in AD transgenic mice.
2. We hypothesized that the suggested beneficial effects of exercise and probiotic treatment have different mechanism, therefore these effects can be summarized and thus reduce the accumulation of Abeta plaques
3. Accordingly to our hypothesis training and/or probiotic treatment will positively modify gut microbiome composition.
4. HIIT will have positive effect on the cognition and will cause a delayed progression of AD.

3. Materials and methods

3.1 Origin of the animals

We have worked with male transgenic mice which were originally obtained from the University of Valencia. Breeding was maintained by the department of Biophysics and Radiation Biology in Semmelweis University under the control of Krisztián Szigeti. With their help we could obtain thirty-two male APP/PS1 transgenic mice (B6C3-Tg (APP^{swe}, PSEN1^{dE9})85Dbo/Mmjax) which were randomly assigned to four groups (n=8 per group) control, exercise, nutrition and combined (exercise and nutrition) group. An additional wild type control group was added to our experiment (n=10). Investigations were performed according to the requirements of “The Guiding Principles for Care and Use of Animals, EU”, and were approved by the Semmelweis University Ethics Committee under the number of PEI/001/2105-6/2014.

3.2 Protocols in the animal house

We have transferred the mice to our animal house when they have reached age 100days. After the transportation has been made, animals had a week of adaptation period. All animals were caged individually and as mice usually lives in colonies we needed to use environmental enrichment. The animals were provided water ad libitum, and they have got 5 grams of classic rodent chow daily. We kept a 12:12 hour light-dark cycle where the light cycle coincided with daytime. All experiments were carried out during the light phase. We have used interval treadmill running and specific probiotic lysate supplementation to test their effects on AD. In our study we have used APP/PS1 transgenic mice in which Abeta plaques are developing from as soon as 6 months, showing cognitive impairment as well. Tau aggregates are not present in our model. Animals were housed individually, because we needed to measure the probiotic intake per animal day by day. Hence for the well being of the animals we needed to use environmental enrichment which consisted of a big cage equipped with a plastic tunnel.

Further than this water was given from a bottle where a rolling ball was in the way of water providing other stimuli to the mice. Running wheels were not implemented because that would interrupt the study design.

All treatments were carried out for 20 weeks. (Figure 6.) Interval treadmill running was applied for the exercise and combined group. Previously all exercising animals were habituated with the motor-driven treadmill (Columbus Inst. Columbus Ohio) and the running speed for 2 weeks. Training was performed four times a week, for 60 minutes. Each training session lasted 10 cycles, each cycle consisted of 4 minutes of high intensity and 2 minutes of low intensity running. Low intensity running speed was permanent during the experiment meaning a speed of 10m/min. While high intensity running speed started at 16m/min and was elevated every third week with 1 m/min until 20m/min was reached. Control and Nutrition group were also habituated with the treadmill and stayed there for 5 minutes/day on a standing treadmill.

Nutrition supplement called Framelim[®] were given 5 times a week 120mg/day for 20 weeks along with the rodent chow. Framelim[®] contains *Bifidobacterium longum* and *Lactobacillus acidophilus* lysate along with B1, B3, B6, B9, B12 vitamins resolved in cod-liver oil. The positive effects of this exact supplement in irritable bowel syndrome and improvements in bowel- and neuropsychiatric symptoms have been reported (122). We have monitored daily food and probiotic lysate uptake, probiotic lysate supplementation did not influence the eating and drinking habits of the animals.

After the 20 week long treatment, animals were exposed to 2 weeks of cognitive testing. We have performed Morris water maze test, Y maze test and open field test. After all the cognitive tests were finished, animals were anaesthetized with an intraperitoneal injection of ketamine (Richter, concentration: 100mg/ml) /xylazine (Produlab Pharma, concentration: 20mg/ml) cocktail in a dose of 0.1 ml/ 10g bodyweight and transcardially perfused with heparinized ice-cold saline.

Brain was removed and measured rapidly, afterward dissected in half along corpus callosum. One hemibrain was postfixed in 4% paraformaldehyde (PFA) for immunohistological staining, the other hemibrain was dissected into 3 parts (frontal, parietal and occipital), and furthermore hippocampus was taken out. All parts were collected, frozen in liquid nitrogen and stored in -80°C until further biochemical analysis.

Fecal samples were also collected for microbiome analysis.

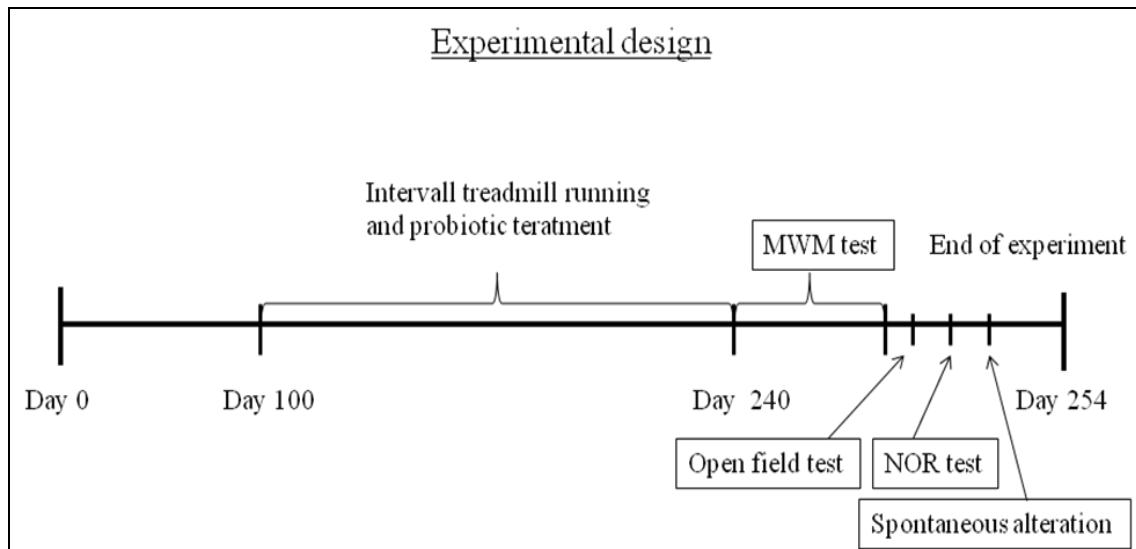


Figure 6. Experimental timeline

Treatments lasted for 20 week from day 100 until day 240. After that cognitive tests were performed.

3.2.1. Cognitive tests

3.2.1.1. Morris water maze test

Morris water maze test was developed by Richard G. Morris in 1984 and since then it is one of the gold standards in behavioral testing. It is used to test spatial memory and learning in rodents. Hippocampus takes part in spatial memory forming, so it is a good way to test hippocampal function in AD transgenic mice (123). In this test, animals can use spatial clues to find the platform, for example posters on the wall, or other objects placed in the testing room.

The test was performed on four consecutive days, each day consisted of four trials. A circular pool with 60 cm in height and 100 cm in diameter were filled with water, and then a six cm wide circular platform was placed in the center of the northwest quadrant of the pool, just one cm below the water surface. Water temperature was maintained between 22° and 23° C throughout all sessions. The test was conducted in darkness so there was no need of water opaquisition. (Figure 7.)

Four starting points were used (north, south, west, or east) and mice were allowed to find the platform for a 60 second period. Each day the order of starting points were mixed. After the 60 second trial animals were allowed to rest on the platform for 30 seconds. This protocol was used for every animal in the experiment. Animal order was randomly assigned. The platform was in the same place on every trial. The latency time to find the hidden platforms was recorded.



Figure 7. MWM test

Morris water maze test in our animal house. Mouse is resting in the hidden platform. 4 starting points are marked with A, B, C and D.

3.2.1.2. Open field test

With this general test it is possible to measure locomotor and exploration activity as well as anxiety. A circular field is used, where outer and inner areas were set in order to measure these spontaneous activities. (Figure 8.) The mouse was placed in the middle of the circle as a starting point, afterwards for a 5-minute time period inner and outer area crossing, rearing, grooming, and latency time was measured and then analyzed.



Figure 8. Open field arena

Open field arena, starting point is in the middle, outer and inner areas are marked.

3.2.1.3. Novel Object Recognition test

This test is performed in the same arena as the Open field test. It can be used the day after the open field test was performed, thus open field test can serve as a habituation test as well. Here the recognition capacity of the mice is evaluated as well as their tendency to restart exploring when they are presented to a novel environment. This trail consists of 2 consecutive test days. On the first day 2 items which are the same size, color and material are placed in the arena. Mice can observe these 2 objects for a certain period of time, and time spent with object observation is recorded. (Figure 9.) On the second test day 1 item is changed to another one which differs in size and material a so called new item. Observation time within old and new object is measured, than analyzed (124).



Figure 9. NOR test arena

Novel object recognition task, part 1. Two equally designed objects are placed in the arena.

3.2.1.4. Spontaneous alternation test

With this test it is possible to investigate the natural behavior of mice, the willingness to explore new areas. In healthy animals it is observed that they are exploring new arms of the maze instead of previously visited ones. Maze arms are set 120 angles apart, starting point is in the middle. Animals are placed individually in the starting point of the maze (Figure 10.) then they could freely explore the arms. We have used a 5-minute trial with a check point at 3 minutes, where latency time and correct alteration was measured. It is counted as a correct alteration when the current entry of the mice differs from the two previous ones, and the 2 previous ones are also different. High rate in alteration refers to sustained cognition. Entry is counted when all 4 limbs are inside the arm as shown in the picture.



Figure 10. Y maze for spontaneous alternation test

Spontaneous alternation test in an Y maze. Mouse is entering one of the arms, all 4 limbs are inside the arm.

3.2.2. Immunohistochemistry

After conservation in 4% PFA for a day brain hemispheres were washed in 0,1% phosphate buffer (PB) in room temperature. Then hemispheres were put into 15% saccharose solution for 4 hours and then placed in 30% saccharose solution overnight in 4° Celsius. Tissue was than embedded in cryoprotectant (Tissue Tek, Sakura Finetek Europe Ref. 4583) over liquid nitrogen.

Brains were than sectioned in a Leica Sliding Microtome (Model SM2000R) to 40 µm coronal sections and stored in PB with sodium-azide.

3.2.2.1. β- amyloid and OGG1 staining

Every sixth free- floating brain section of groups was immunostained for amyloid plaques (6E10, Anti-β- amyloid, 1-16 antibody, BioLegend #803015). Beta-amyloid was detected with an affinity-purified antiserum from a mouse immunized with human antigen. This antibody (1:15000 for 3,3'-diaminobenzidine) was applied overnight at room temperature followed by incubation of the sections in biotinylated antimouse secondary antibody 1 h (1:1000 Vector Laboratories #BA 2000) and then in avidin-biotin-peroxidase complex (1:500, VECTASTAIN Elite ABC-Peroxidase Kit, PK-6100) for 1h. Subsequently, sections were treated with 0.06% diaminobenzidine (DAB, sigma) 0.08% nickel (II) sulfate and 0.003% H₂O₂ in Tris-hydrochloride buffer (0.1M, pH8.0) for 2.5 min, mounted and coverslipped.

DNA repair enzyme (Anti-OGG1 Abcam #ab22766) was visualized using the same method described above. The antibody was used in a dilution of 1:2000. A biotinylated antirabbit secondary antibody was used (Jackson ImmunoResearch #711-065-152)

3.2.2.2. Double labeling 6E10 and Iba1

Brain sections of animals were processed for double labeling with 6E10 and Iba1. Every sixth free-floating section was first stained for Iba1 (AIF/Iba1 Novus biologicals # NB100-1028) by using FITC-tyramide amplification fluorescent

immunocytochemistry. This antiserum (1:2500) was applied overnight at room temperature, followed by incubation of the sections in biotinylated antigoat secondary antibody (1:1000, Vector Laboratories# BA 9500), afterwards in ABC complex (1:500; Vector Laboratories) for 1h. Then sections were subsequently incubated with FITC-tyramide (1:8000, Sigma) and H₂O₂ in Tris hydrochloride buffer (0.1 M, pH 8.0) for 8 min. Sections were then incubated overnight in 6E10 (1:7000; BioLegend #803015) at room temperature. Following application of the primary antibody, sections were incubated in Alexa Fluor 594 antimouse secondary antibody (Invitrogen #A21203) for 2h. After washes, sections were mounted and coverslipped.

3.2.2.3. Microscopy and image processing

An Olympus BX60 light microscope equipped with fluorescent epi-illumination and a dark-field condenser were used to examine the sections. Images were captured at 2048 × 2048 pixel resolution with a SPOT Xplorer digital CCD camera (Diagnostic Instruments, Sterling Heights, MI) using 10-20 × objectives. Images were adjusted using the “levels” and “sharpness” commands in Adobe Photoshop CS5.1. Full resolution of the images was maintained until the final versions, which were adjusted to a resolution of 300 dpi. Images were analyzed with ImageJ Software version 1.48v.

3.2.3. Western blotting

Hippocampi were homogenized in a buffer containing: 137mM NaCl, 20mM Tris-HCl pH8.0, 2% NP40, 10% glycerol and protease inhibitors (PMSF, aprotinin, orthovanadate). Protein levels were measured using Bradford Protein assay (Bio- Rad, # 500-0006).

Proteins were electrophoresed on 8-12% v/v polyacrylamide SDS-PAGE gels, and then were transferred onto PVDF membranes. The membranes were blocked in 5% milk powder containing Tris-buffered saline with Tween 20 (TBS-T). After blocking, membranes were incubated overnight at 4° C with the following antibodies BDNF (1:500, Santa Cruz Biotechnology # sc-546), PSMA6 (1:1000, Cell signaling # 2459).

IDE and Neprilysin. Following incubation with primary antibodies, membranes were washed 3x20 minutes with TBS-T and incubated with horseradish peroxidase (HRP)-conjugated secondary antibodies (1:10000) at 4° C for 2 hours. Afterwards membranes were repeatedly washed with TBS-T. Then membranes were incubated with HRP reagent (Super Signal West Pico Chemiluminescent Substrate, Thermo Scientific #34080) for 5 minutes, and protein bands were visualized on X-ray films. Bands were quantified using ImageJ software version 1.48v and normalized to alfa- tubulin.

3.2.4. Library preparation and identification of prokaryotic species

The DNA from stool samples was isolated by QIAmp Fast DNA stool mini kit (Quiagen). Fragment libraries were constructed from purified DNA using NEBNext Fast DNA Fragmentation & Library Prep Set for Ion Torrent (New England Biolabs) according to manufacturer's instructions. Briefly, DNA was enzymatically digested, and the fragments were end-repaired. Ion Xpress Barcode Adaptors (Life Technologies) were then ligated and the template fragments size-selected using Agencourt AMPure XP magnetic beads (Beckman Coulter). Adaptor ligated fragments were then PCR amplified, cleaned up using AMPure beads, quality checked on D1000 ScreenTape and Reagents using TapeStation instrument (Agilent) and finally quantitated using Ion Library TaqMan Quantitation Kit (Life Technologies). The library templates were prepared for sequencing using the Life Technologies Ion OneTouch protocols and reagents. Briefly, library fragments were clonally amplified onto Ion Sphere Particles (ISPs) through emulsion PCR and then enriched for template-positive ISPs. More specifically, PGM emulsion PCR reactions utilized the Ion PGM Hi-Q OT2 Kit (Life Technologies), and as specified in the accompanying protocol, emulsions and amplification were generated using the Ion OneTouch System (Life Technologies). Enrichment was completed by selectively binding the ISPs containing amplified library fragments to streptavidin coated magnetic beads, removing empty ISPs through washing steps, and denaturing the library strands to allow for collection of the template-positive ISPs. For all reactions, these steps were accomplished using the Life Technologies ES module of the Ion OneTouch System. Template-positive ISPs were deposited onto the

Ion 318 chips (Life Technologies); finally, sequencing was performed with the Ion PGM Hi-Q view OT2 Kit (Life Technologies).

3.3 Statistics

In the beginning of the experiment transgenic animals were randomly assigned to the following groups: Control-APP/PS1^{TG}-C, exercise- APP/PS1^{TG}-Ex, nutrition-APP/PS1^{TG}-Pr and combined (exercise and nutrition) group- APP/PS1^{TG}-Ex-Pr. An additional wild type control group WT was added to the experiment. Each transgenic group consisted of 8 animals while WT group consisted of 10 animals.

For statistical analysis, except for microbiome samples, we used GraphPad Prism 5 Software. For the evaluation of physiological and biochemical data, we first performed Shapiro-Wilk normality test of all dependent variables. Based on the result we performed one-way analysis of variance (ANOVA) followed by Tukey's post hoc test or Kruskal-Wallis ANOVA with Dunn's post hoc test. Significance level was set at $p < 0.05$.

3.3.1. Microbial analytical methods

Bacterial genome annotation was carried out by uploading the FASTQ data files to the automated web-based metagenomics analysis server–MG-RAST version 3.6. (125). MG-RAST takes FASTQ data files as input, identifies open reading frames that are likely to be genes, and uses a series of subsystem techniques (the 'ST' in RAST) to compare these with a sophisticated database of genes and RNA sequences, producing a high-quality annotation of the assembly. The data files, annotated based on the RefSeq database were downloaded for further analysis. The MG-RAST ids of the datasets are the following: Wt (healthy control): mgm4672670.3; APP/PS1^{TG}: mgm4673335.3; APP/PS1^{TG} -Ex: mgm4682164.3; APP/PS1^{TG} -ExPr: mgm4682165.3; APP/PS1^{TG} -Pr: mgm4682167.3.

3.3.2. Bioinformatics analysis of the microbiome

The data were filtered based on the following criteria of annotation quality: minimum alignment length: 30 base pairs; minimum percentage of identity: 60%; maximal E-value: 10^{-5} . Then, the annotated reads of each sample were permuted in random order, and were divided into 10 non-overlapping subsets, containing 10% of the original data. This process was performed via Python script. The generated populations were used for calculating the relative abundances and standard deviations of selected microbial groups. The results were visualised in bar graphs. The significance of differences between the groups was tested with two-sample Kolmogorov-Smirnov test with $p < 0.001$ significance threshold. These analyses were carried out in MATLAB.

4. Results

4.1. Results from the animal experiments

4.1.1. Animal weight change during the experiment

We checked the body mass of the animals weekly during the experiment. We did not observe any major changes in animal weight, proving that none of the treatments meant serious stress factor to the animals. All animals gained a little weight by the end of experiment. (Figure 11.) Interesting that lowest weight was observed in the APP/PS1^{TG}-ExPr group nevertheless they received the probiotic lysate supplement which consisted of cod liver oil, which supposed to give them extra energy.

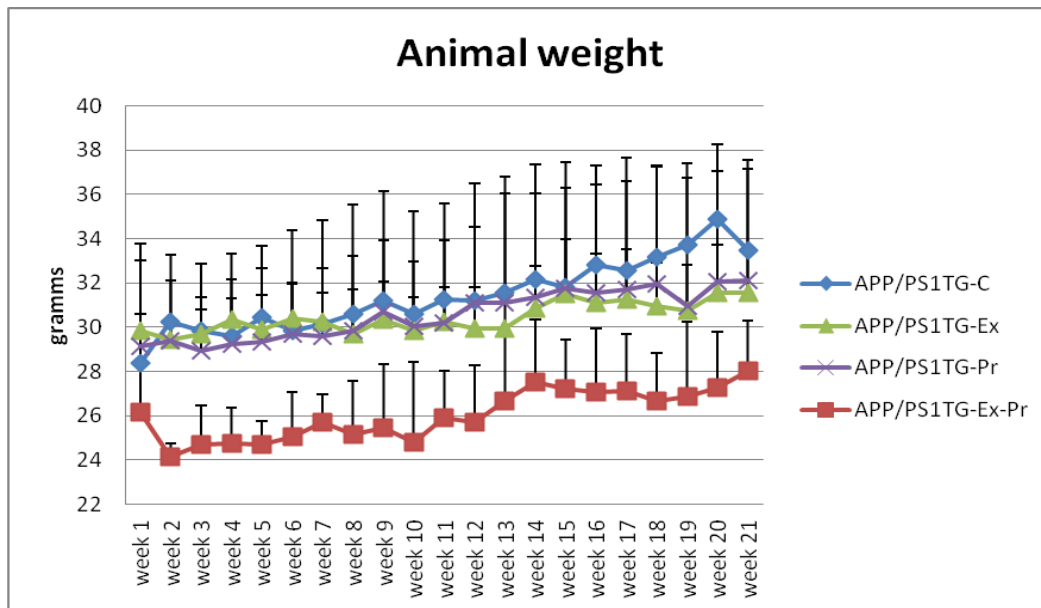


Figure 11. Body mass change during experimental period

Body mass change during 20 weeks of experimental period, plus 1 week of test period.

N=8, Results are expressed as mean+ SD No significant differences were measured among study groups. There were no significant differences in weight gain or loss in any of the groups.

4.1.2 Cognitive test results

In cognitive test we used wild type mice (N= 10) to evaluate the performance of transgenic mice compared to this group. Also, we have analyzed the results only between the transgenic groups.

Spatial memory was tested with MWM test, where wild type mice did not outperform all transgenic groups. Interestingly APP/PS1^{TG}-Ex-Pr group performed the best among all study groups, indicating a well-preserved brain function. (Figure 12.)

If we investigate only the performance of transgenic groups, we will find significant differences among the groups. APP/PS1^{TG}-Ex-Pr group outperformed the other transgenic groups in the second third and fourth day.

On the second day APP/PS1^{TG}-Ex-Pr group outperformed the APP/PS1^{TG}-C group.

On the third day APP/PS1^{TG}-Ex-Pr group outperformed the APP/PS1^{TG}-Ex group.

On the fourth day APP/PS1^{TG}-Ex-Pr group outperformed the APP/PS1^{TG}-Ex and APP/PS1^{TG}-Pr group.

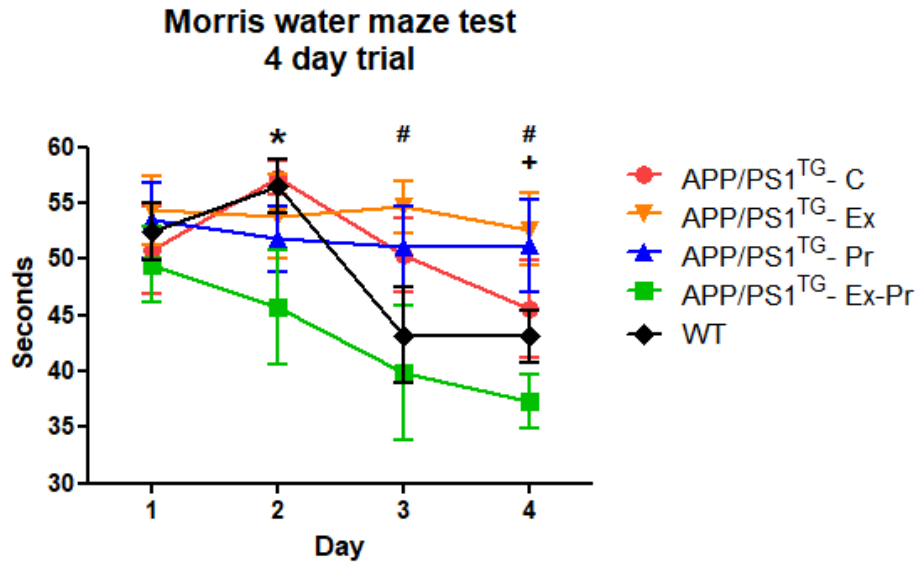


Figure 12. MWM test results including WT

Morris water maze test results. (N=8 in APP/PS1^{TG} groups and N=10 in WT.)

Results are expressed as mean \pm SD. Legend showing significant differences ($p < 0.05$)

* C vs ExPr, # Ex vs ExPr, + Pr vs ExPr

No significant differences were observed between the transgenic groups and wild type group. Combined treatment receiving group APP/PS1^{TG}-Ex-Pr performed better than wild type group, albeit not significantly.

APP/PS1^{TG}-Ex-Pr animals performed significantly better in the MWM, especially on the 4th testing day. They outperformed both treated groups, they have found the hidden platform in a shorter period of time, indicating that in AD exercise or dietary supplements cannot change radically hippocampal function, while combined treatment can have a spatial memory reserving function.

On the open field testing we have found no significant differences in exploratory activity among the study groups. All groups entered in the inner and in the outer areas as well, which means that none of the animals suffered from anxiety, they were all ready to explore. There were no significant differences between inner nor in outer crossing. (Figure 13.A.)

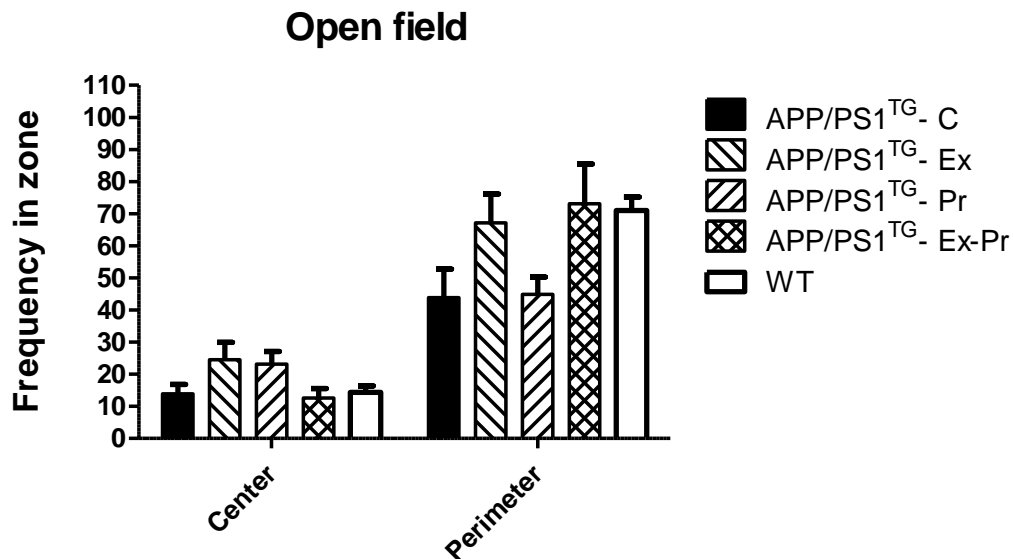


Figure 13.A. Open field test exploratory activity

Open field test results. (N=8 in APP/PS1^{TG} groups and N=10 in WT.)

Results are expressed as mean \pm SD. No significant differences were observed among the study groups, indicating that animals were not stressed. Exercise did not alter their activity behavior.

In latency times, we have found significant differences between the wild type and APP/PS1^{TG}-C group. Suggesting the possibility, that even though there were no significant differences in exploratory activity there might be some sign of anxiety in transgenic groups. If we only compare the transgenic groups among each other we cannot find any significant differences between them. (Figure 13.B.)

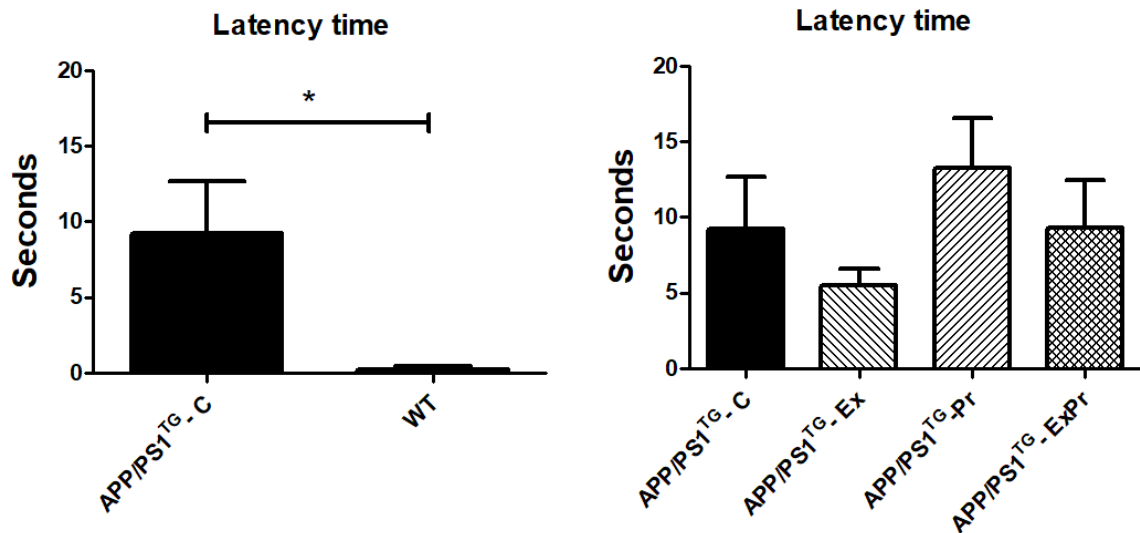


Figure 13.B. Latency time in OFT

Differences in latency time in the open field test. (N=8 in APP/PS1^{TG} groups and N=10 in WT.) Significant differences are shown with *, significance level is $p < 0.05$ Results are expressed as mean \pm SD.

Wild type animals almost started to explore the field right after placing them in the arena while APP/PS1^{TG}-C animals needed significantly more time, almost 10 seconds to start the test. Among transgenic groups probiotic lysate treated group stayed still for the longest time.

If we observe total exploration activity, we could not find any significant differences. Showing that AD animals do not have altered behavior in case of exploration. None of the treatments affected their behavior inversely. (Figure 13.C.)

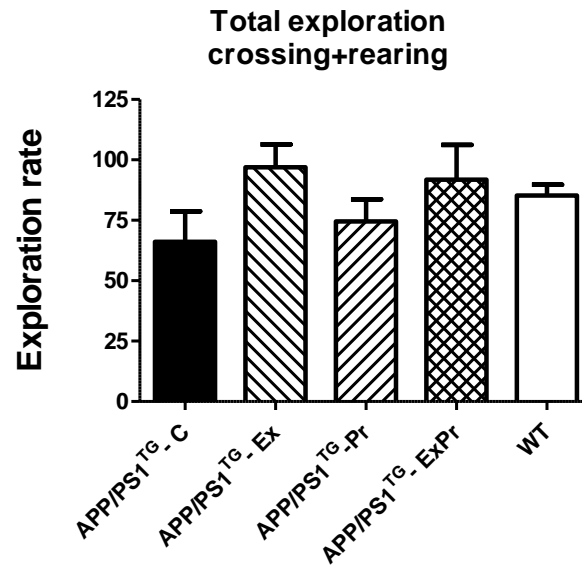


Figure 13.C. Total exploration in OFT

N=8 in APP/PS1^{TG} groups and N=10 in WT. Results are expressed as mean ± SD

Open field total exploration activity shows no significant differences among study groups. Total exploration means the total number of crossing activity plus the rearing number.

Novel object recognition task. Open field test needs to be performed previous to the NOR test, because animals should explore the objects in the arena instead of the new environment. Mice recognize the old object as a familiar one, thus spending more time discovering the new one. APP/PS1^{TG}-Ex mice significantly spent less time with the new object compared to the group APP/PS1^{TG}-C (Figure 14.)

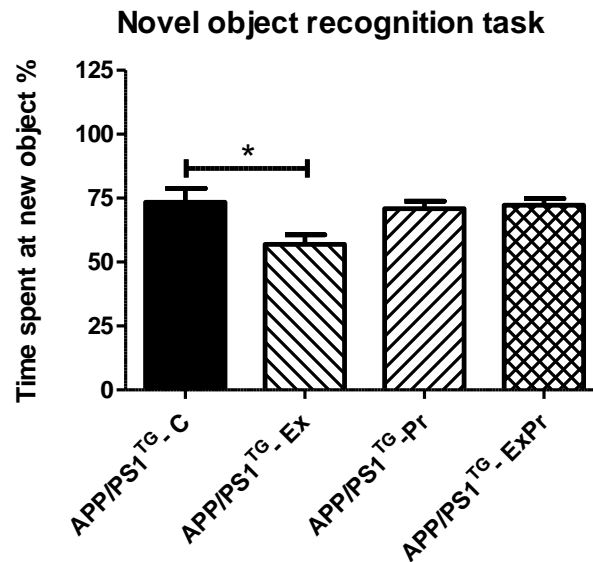


Figure 14. NOR test

Novel object recognition task, second observation with new object in the arena. Time spent with the new object shows that animal is able to remember that this object was not there for the first investigation. Mice innate instinct is to spend more time with a new, non familiar object. Results are shown in %. % = time spent with the new object/ all the time spent with both objects *100. (N=8) Results are expressed as mean \pm SD

Significance level is $p < 0.05$, significant differences were marked with *.

Spontaneous alternation is a good method to measure cognitive deficits in transgenic mouse models. As data with WT mice shows well, AD has serious effects on cognition. APP/PS1^{TG}-C animals performed significantly poorer than WT animals if we measure the % of good entries. If transgenic groups are compared we can find no significant differences in the % of good alterations. (Figure 15A, B)

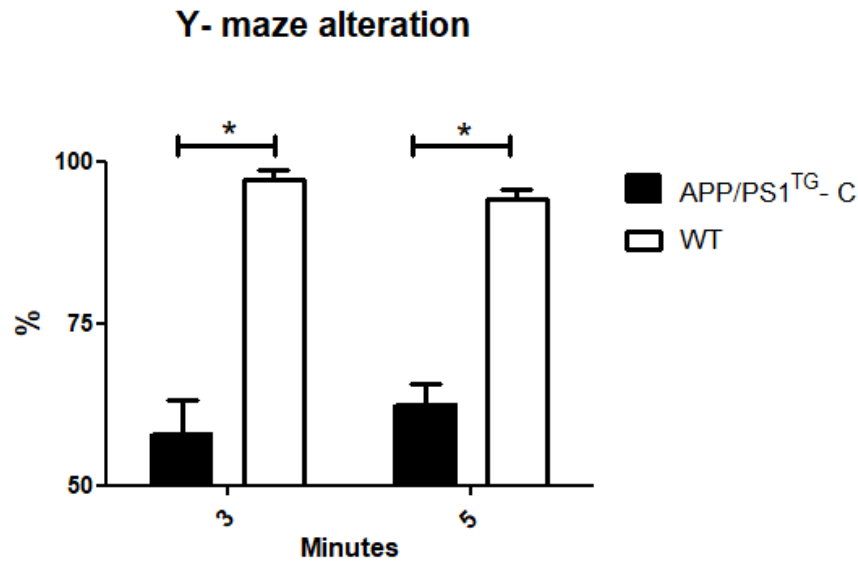


Figure 15.A. Spontaneous alternation test, APP/PS1^{TG}-C vs WT (N=8 in APP/PS1^{TG} groups and N=10 in WT.) Results are expressed as mean \pm SD

Significant differences were marked with *. $p < 0.05$

In spontaneous alternation test WT mice outperforms significantly the transgenic control group, showing better cognitive function in working memory and in spatial orientation.

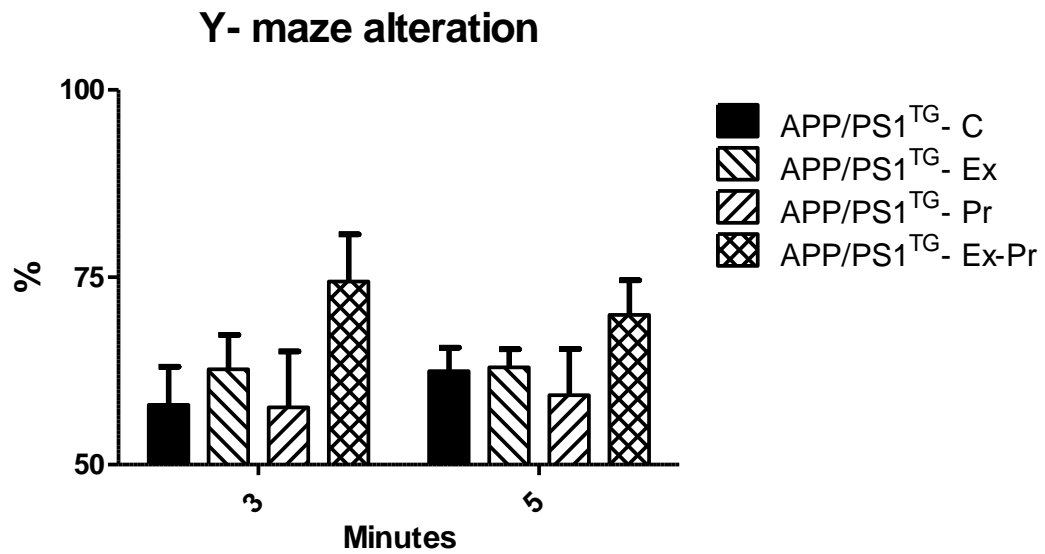


Figure 15.B. Spontaneous alternation test, transgenic groups
(N=8) Results are expressed as mean \pm SD

If we check the performance of transgenic groups no significant differences can be found. None of the groups performed significantly better than the others, animals showed similar exploratory activity and similar correct alteration percentage

If we only count the number of entries, we can find no differences. Data shows that there is no difference in the activity of animals. Neither of the treatments nor the disease had an impact on the mobility of animals. (Figure 16.)

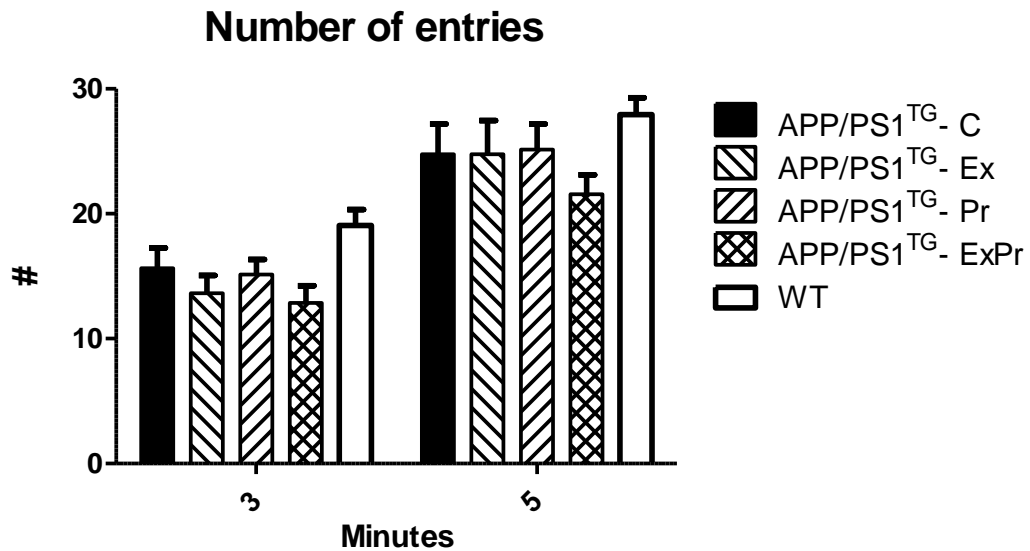


Figure 16. Spontaneous alternation, number of entries (N=8 in APP/PS1^{TG} groups and N=10 in WT.) Results are expressed as mean \pm SD. No significant difference was shown among the groups, all groups were similarly active. Number of entries did not differ significantly.

4.2. Results from brain tissue investigation

Amyloid plaque staining with 6E10 antibody shows that amyloid plaque deposition was most abundant in the hippocampal region as it was expected in this mouse strain. (Figure 17.) Most plaques were observed in the APP/PS1^{TG}-C group. Compared to APP/PS1^{TG}-C group (31.7 ± 9.0) and APP/PS1^{TG}-ExPr group (29.1 ± 6.7) in the APP/PS1^{TG}-Ex group (13.9 ± 6.7) significantly lower level of plaques were counted. Interestingly we can notice that all treated groups had a significantly lower level of area covered by plaques compared to APP/PS1^{TG}-C group.

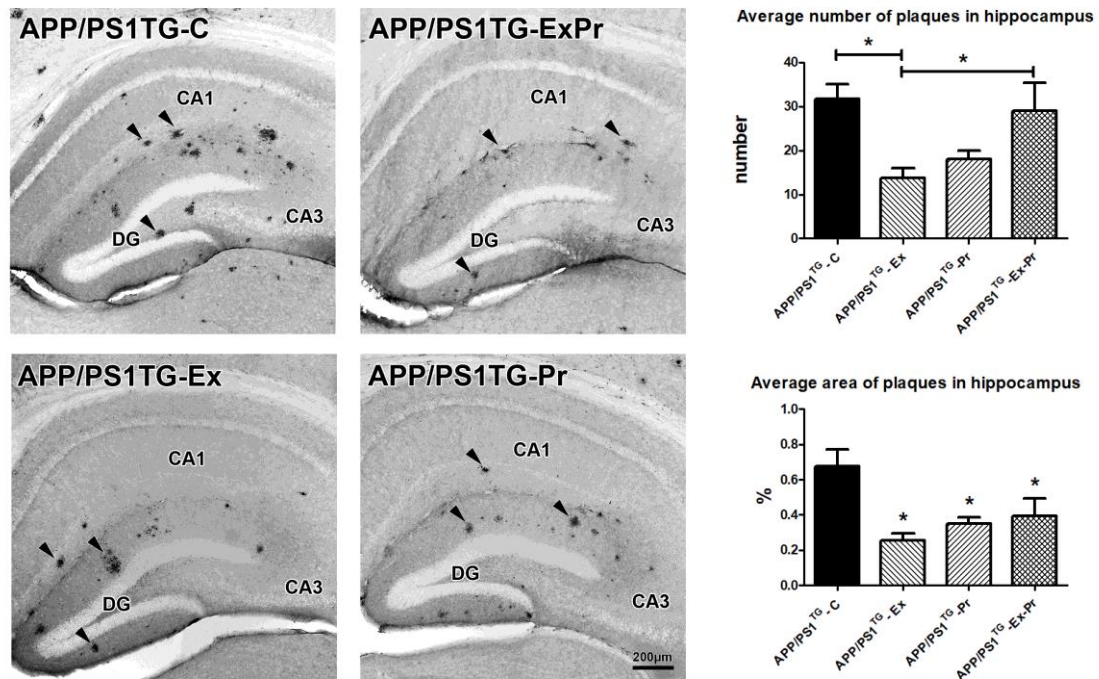


Figure 17. Amyloid plaque depositions in hippocampus

Histochemical measurement shows that exercise not only reduced the plaque number, but it also reduced the area covered by the plaques compared to APP/PS1^{TG}-C groups.

Amyloid plaque area was decreased in all treated groups.

(N=8) Results are expressed as mean ± SD * $p < 0.05$

We have measured the microglia number in the hippocampal area as well. (Figure 18.) In one hand we did not find any differences in the total number of microglia among the groups. On the other hand area covered by microglia was significantly higher in APP/PS1^{TG}-ExPr group (8.11 ± 1.3) than in the APP/PS1^{TG}-C group (5.36 ± 2.4)

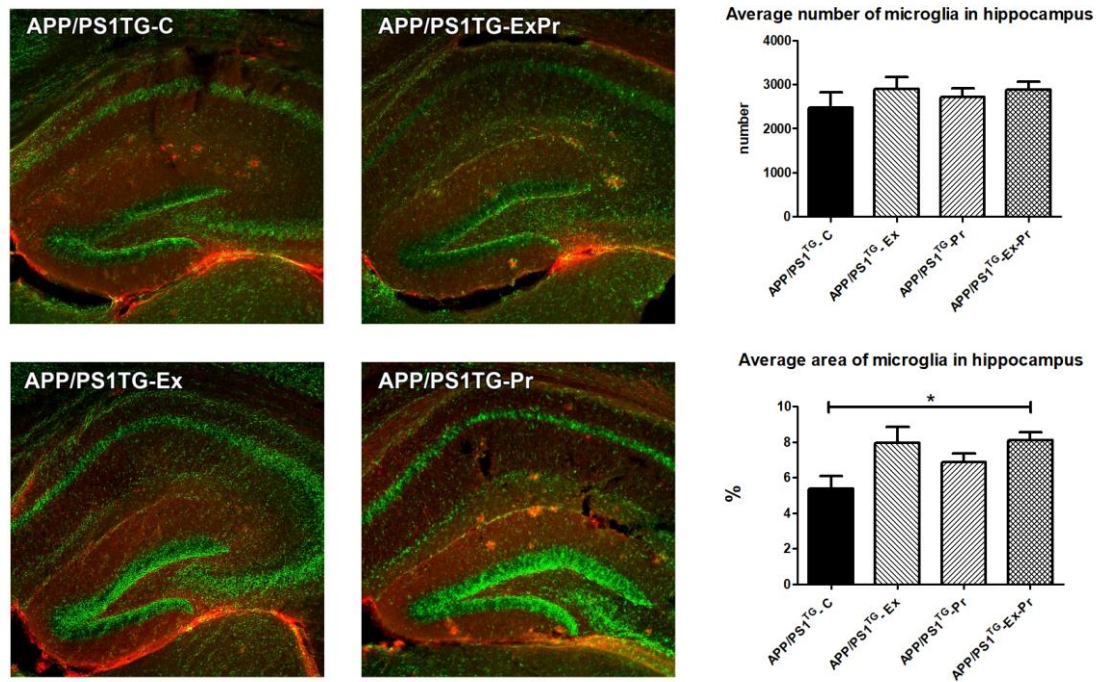


Figure 18. Microglia number and area in the hippocampus

Area covered by microglia is increased in all treated groups compared to APP/PS1^{TG}-C group. Furthermore, it can be observed that microglia (marked with green) is accumulated around the amyloid plaques (red), which could predict elevated microglial reparational functions.

(N=8) Results are expressed as mean \pm SD * p<0.05

OGG1 levels are decreased in AD. Exercise can raise the OGG1 levels in the brain. We have measured if exercise can modify the levels of OGG1 (Figure 19.) It turned out that OGG1 levels were the highest in the APP/PS1^{TG}-Ex group (7951 ± 4085), it was significantly higher than in other treated groups APP/PS1^{TG}-Pr (2947 ± 2222) and APP/PS1^{TG}ExPr (3358 ± 1127). Average area demonstrates the same pattern.

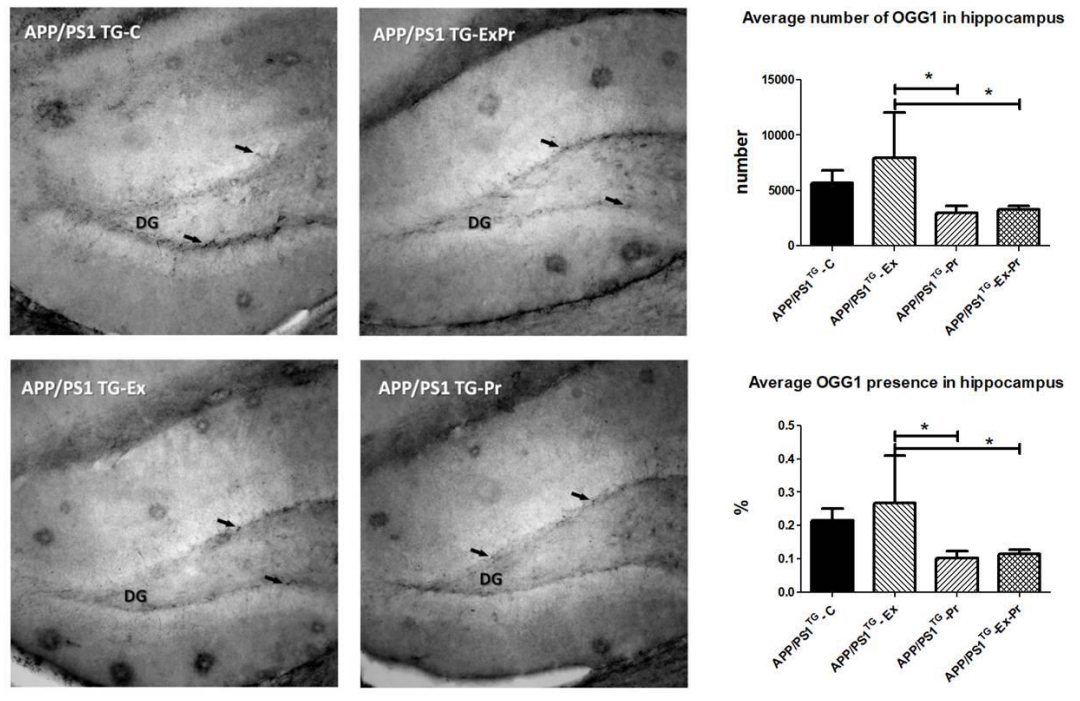


Figure 19. OGG1 levels in hippocampus

While exercise increased OGG1 level, probiotic lysate seems to prevent this effect. Moreover, probiotic lysate reduced OGG1 level below the level it was observed in the APP/PS1^{TG}-C group.

(N=8) Results are expressed as mean \pm SD * $p < 0.05$

Western blot analysis did not result in any significant differences which can be a proof or denial to our findings, thus these data is not represented here.

We have performed ELISA measurement regarding A β 40 and A β 42 levels. We did not find any significant differences either.

4.3. Results from microbiome analysis

DNA from fecal samples of mice were analyzed to measure the effects of treatments. Additionally WT mice were used to investigate if APP/PS1 transgenic mice have an altered microbiome or not. In general, at genus level *Firmicutes* and *Bacteroidetes* were the most abundant, while Proteobacteria, Actinobacteria, Fusobacteria and Verrucomicrobia were less present in the mice microbiome samples. (Figure 20.) The Firmicutes/Bacteroidetes ratio was the lowest in APP/PS1^{TG}-Pr, and the highest in WT group.

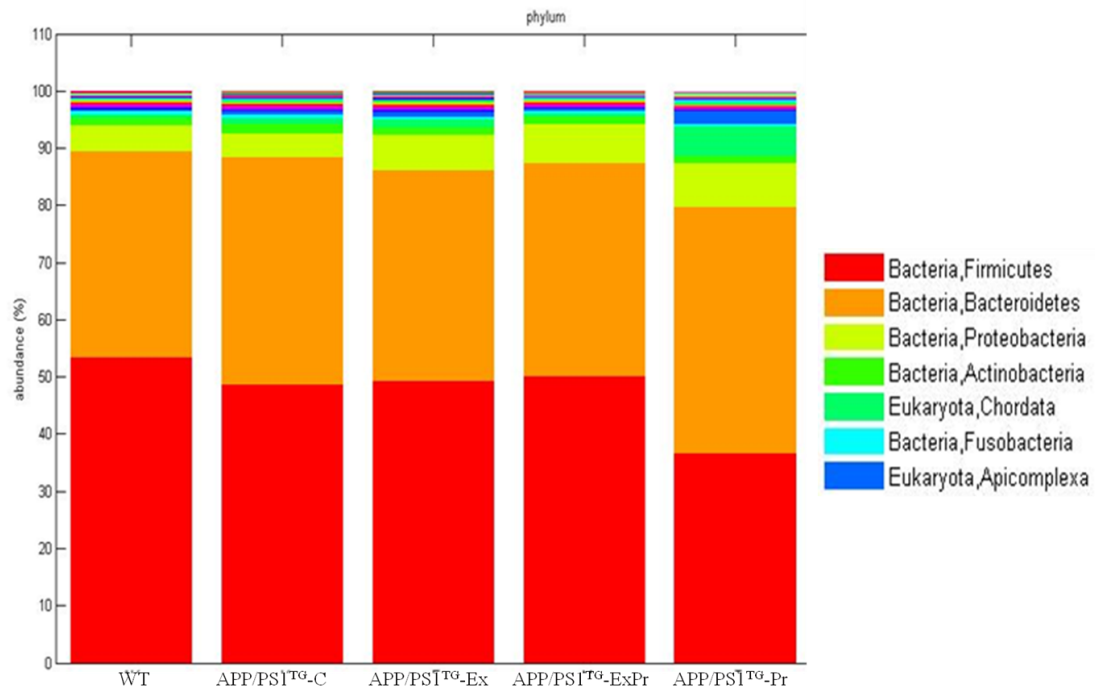


Figure 20. Microbiome distribution in phylum level

Most abundant presence was measured in Firmicutes and Bacteroidetes phyla among all study groups.

Differences in microbiome composition between WT mice and APP/PS1^{TG}-C mice are more outstanding if we investigate them at genus level. (Figure 21 A, B.) *Bacteroides spp.* was significantly lower while *Prevotella spp.* was significantly higher in WT group compared to APP/PS1^{TG}-C group (Figure 21.A.)

Butyrate producing genres such as *Clostridium spp.* *Eubacterium spp.* and *Roseburia spp.* were found in elevated levels in WT group suggesting a more preserved mucin layer in the gut. (Figure 21.B.)

If genus differences are compared between the transgenic groups, it can be seen that exercise had a positive effect on butyrate producing genres while probiotic receiving group had the lowest levels of these bacteria. (Figure 21.C.)

The probiotic lysate supplement contained Omega 3 fatty acids which should elevate the levels of *Lactobacillus spp* (Figure 21.D.) and should decrease the levels of *Clostridium spp.* (Figure 21.C.).

Strains with effects on mucin production

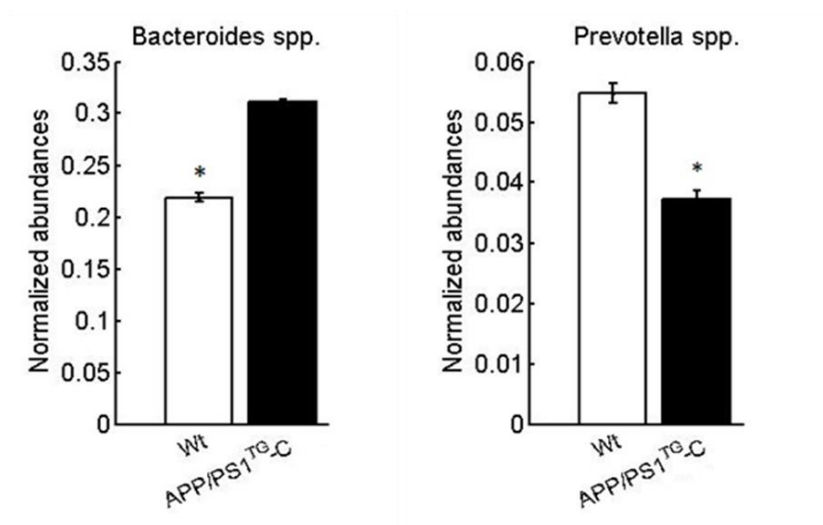


Figure 21.A. Strains with effects on mucin production WT vs APP/PS1^{TG}-C
 Low levels of *Bacteroides spp.* and high levels of *Prevotella spp.* in WT group suggest the presence of physiological mucin generation on the epithelial layer in the GIT. *Bacteroides spp.* is a succinate producer which can have a negative effect on mucin layer thickness.

(N=8). * $p < 0.001$. Results are expressed as mean \pm SD, from the two-sample Kolmogorov-Smirnov test.

Butyrate producer strains

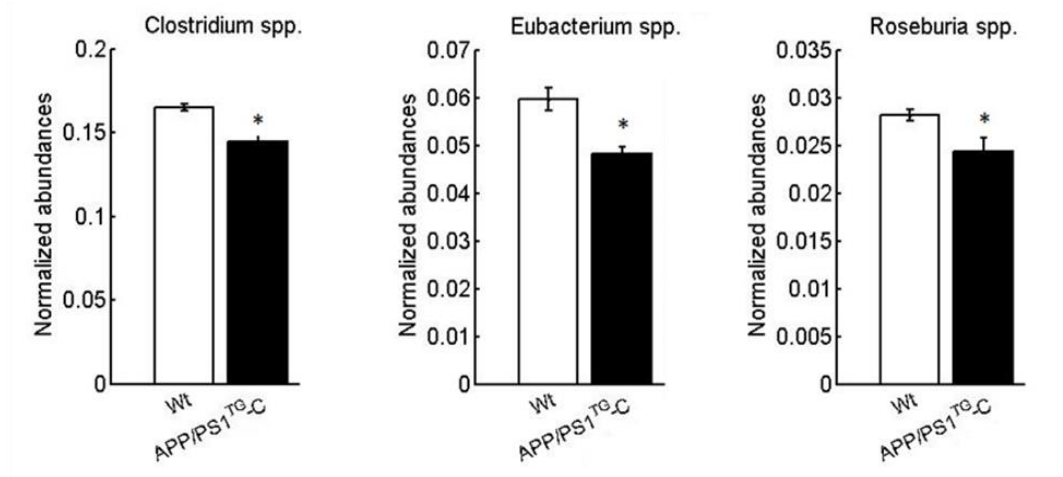


Figure 21.B. Butyrate producer strains WT vs APP/PS1^{TG}-C

Butyrate producer genres were also present in a significantly higher level in WT than in APP/PS1^{TG}-C group. Data suggest that AD has a negative effect on the microbiome composition suggesting the presence of leaky gut.

(N=8). * $p < 0.001$. Results are expressed as mean \pm SD, from the two-sample Kolmogorov-Smirnov test.

Butyrate producer strains

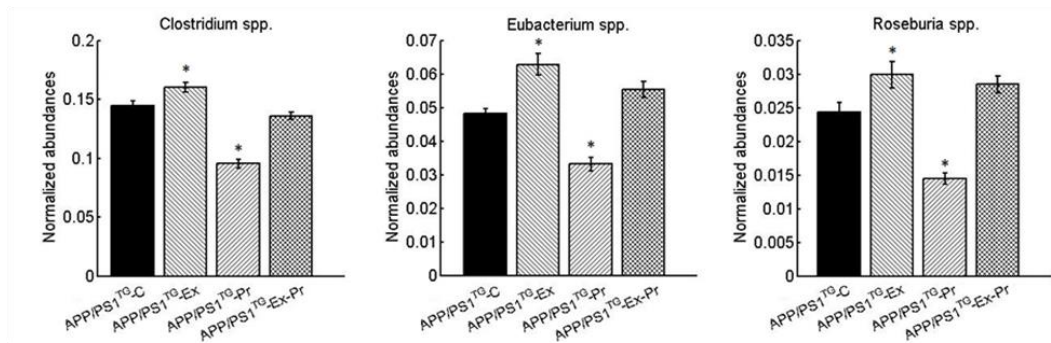


Figure 21.C. Butyrate producer strains, transgenic groups

There were significant differences among APP/PS1^{TG}-Ex and APP/PS1^{TG}-Pr groups in the abundance of butyrate producing genres. Proving evidence that exercise has a beneficial effect on butyrate producing bacteria.

(N=8). * $p < 0.001$. Results are expressed as mean \pm SD, from the two-sample Kolmogorov-Smirnov test.

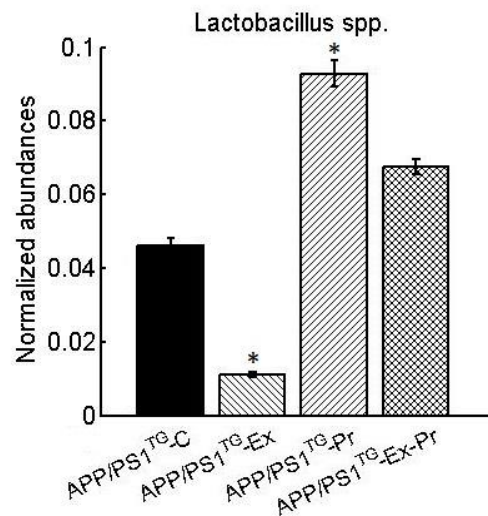


Figure 21.D. Effect of probiotic lysate on *Lactobacillus spp.*

Exercise decreased the abundance of *Lactobacillus spp.* while probiotic lysate elevated it. It seems that probiotic lysate is effective and enough to elevate the abundance of *Lactobacillus spp.* in the GIT, there is no necessary need for probiotics.

(N=8). * $p < 0.001$. Results are expressed as mean \pm SD, from the two-sample Kolmogorov-Smirnov test.

At the species level we can observe other interesting differences.

Comparing WT and APP/PS1^{TG}-C groups we can observe that *Bacteroides* species *Bacteroides thetaiotaomicron* and *Bacteroides fragilis* are present in a significantly lower level in WT group, again suggesting a presence of a better conserved epithelial layer. Elevated levels of butyrate producer *Butyrivibrio proteoclastus* is in accordance with the previous suggestion. (Figure 22.A.)

Levels of *Bacteroides thetaiotaomicron* were significantly higher in APP/PS1^{TG}-Pr compared to APP/PS1^{TG}-ExPr group, this result correlates well with results from MWM test. Elevated levels of this bacteria can suggest the presence of leaky gut syndrome because they produce acetate and succinate as the end product of glucose and lactose fermentation. Both products are responsible for decreased mucin production. Probiotic lysate elevated the levels of *Bacteroides fragilis*, interestingly exercise could modify this effect. Exercise receiving groups APP/PS1^{TG}-Ex and APP/PS1^{TG}-ExPr have a significantly lower number from this bacteria than APP/PS1^{TG}-Pr group. *Lactobacillus reuteri* is a well known B12 vitamin producer, which is essential for optimal brain function and it has an elevated level in APP/PS1^{TG}-Pr and APP/PS1^{TG}-ExPr group. *Lactobacillus johnsoni* which is found to generate excessive H₂O₂ has significantly lower levels in APP/PS1^{TG}-Ex group compared to APP/PS1^{TG}-C proving an evidence for the positive role of exercise against oxidative stress. Butyrate producing bacteria had different abundance levels among the groups. These bacteria are *Butyrivibrio proteoclastus*, *Marvinbryantia formatexigens*, *Roseburia intestinalis* and *Roseburia inulinivorans* all of this had an increased level in exercise receiving groups APP/PS1^{TG}-Ex and APP/PS1^{TG}-ExPr compared to only probiotic lysate receiving group APP/PS1^{TG}-Pr. (Figure 22.B, C.)

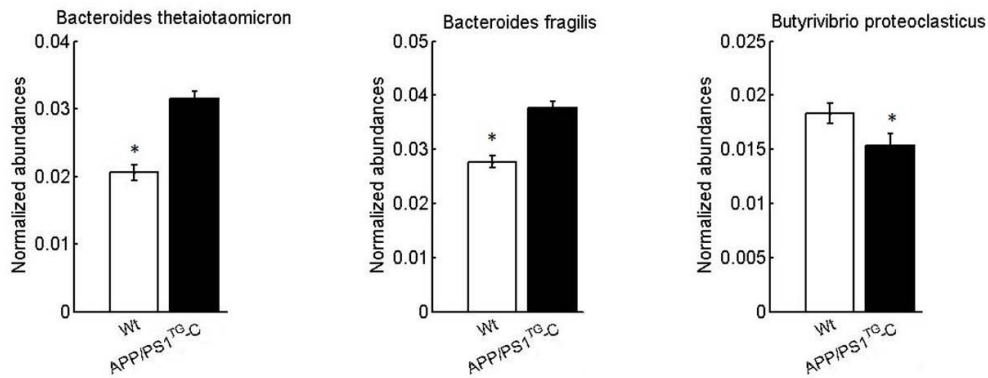


Figure 22.A. Species differences among WT and APP/PS1^{TG}-C

Low levels of *Bacteroides thetaiotaomicron* and *Bacteroides fragilis* and elevated level of butyrate producer *Butyrivibrio proteoclasticus* in WT group suggest a better state of the intestinal mucin layer.

(N=8). * $p < 0.001$. Results are expressed as mean \pm SD, from the two-sample Kolmogorov-Smirnov test.

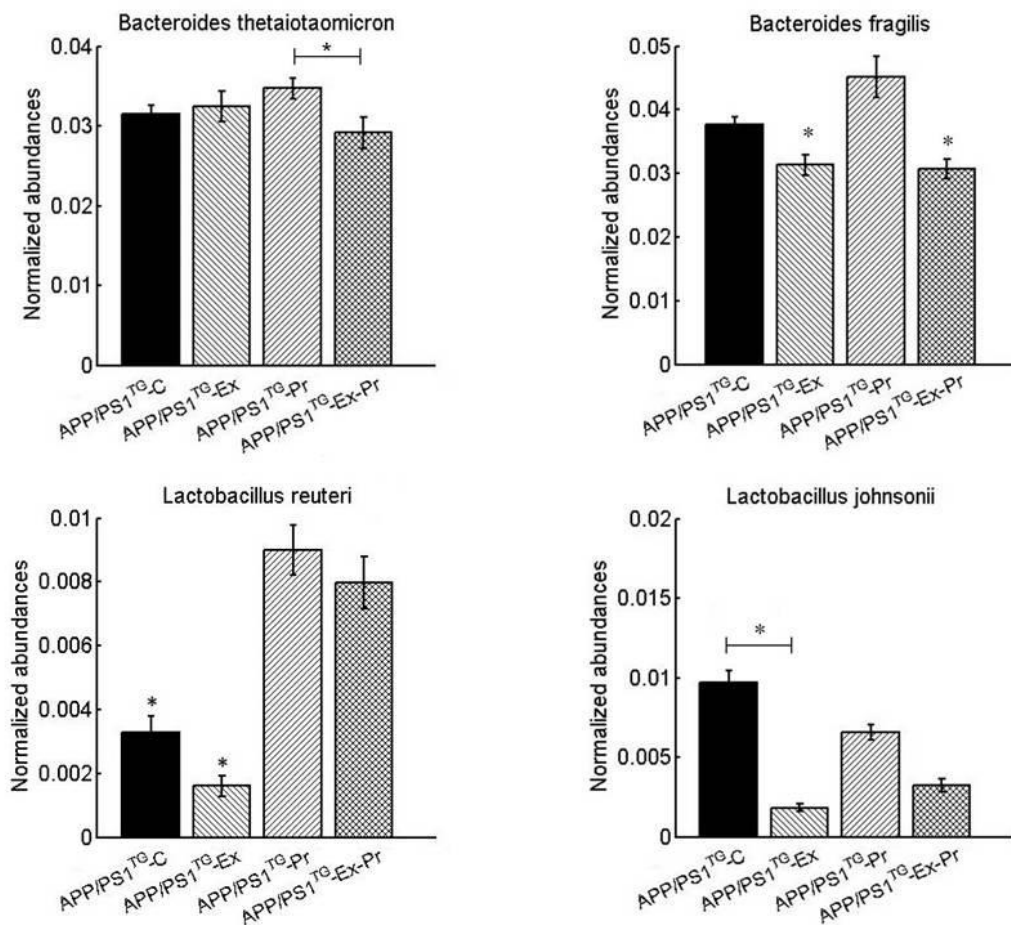


Figure 22. B. Species differences among transgenic groups I.

It can be observed that probiotic lysate elevated the levels of *Bacteroides* species which are responsible for acetate production and *Lactobacillus reuteri* which is responsible for B12 vitamin production. These effects are controversial while acetate may be responsible for the development of leaky gut, B12 vitamin is essential for proper brain function.

(N=8). * $p < 0.001$. Results are expressed as mean \pm SD, from the two-sample Kolmogorov-Smirnov test.

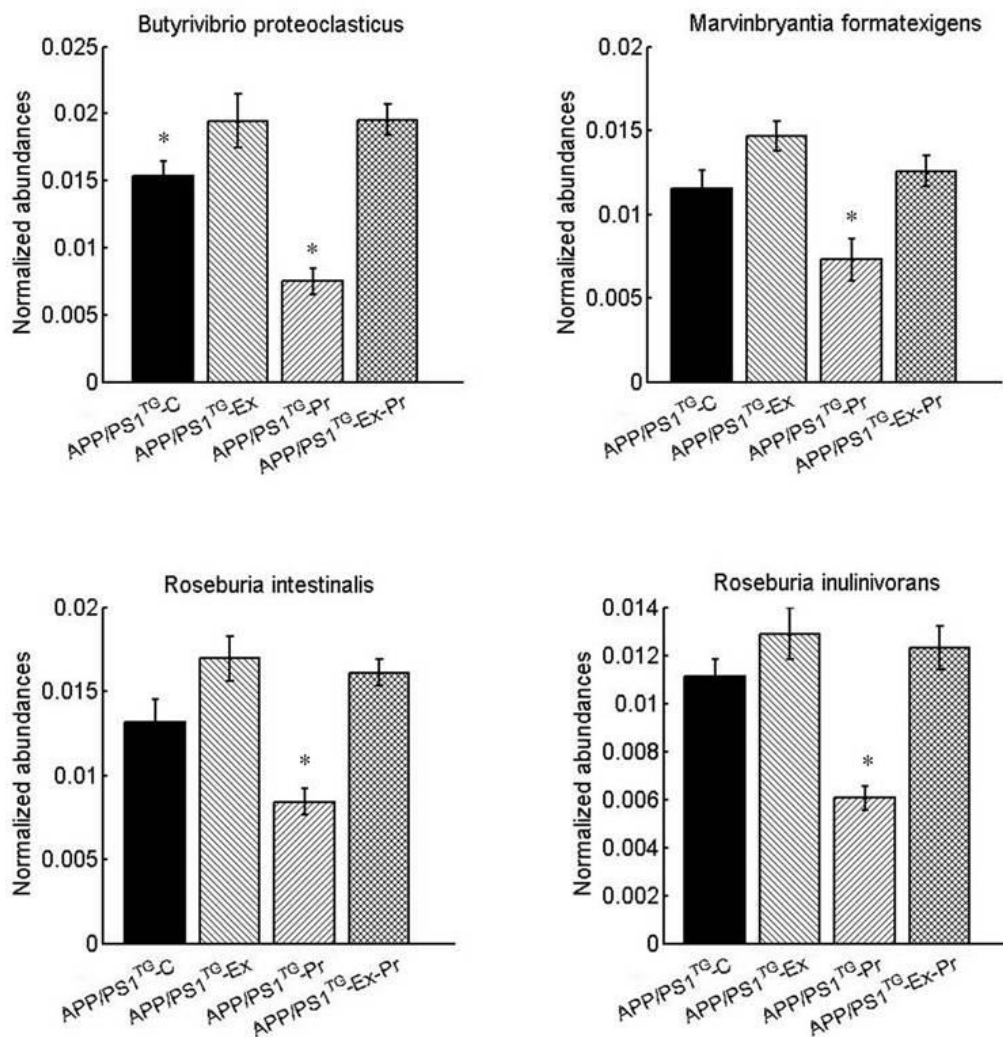


Figure 22. C. Species differences among transgenic groups II.

Exercise receiving groups had an elevated number of butyrate producing species suggesting a more preserved mucin layer in the gut.

(N=8). * $p < 0.001$. Results are expressed as mean \pm SD, from the two-sample Kolmogorov-Smirnov test.

5. Discussion

Regular physical activity (56-58) and dietary habits (120, 121) were in the scope for AD prevention in the last years. To approve this assumption, we studied APP/PS1 transgenic male mice, and treated them for 20 weeks with interval treadmill running and/or probiotic lysate supplementation. Treatments were started before amyloid plaque formation begins in this model. This way the preventive effects of the treatments could be observed.

We were interested if HIIT can have the same positive effects on cognition in animal models as it was proven before with MICT (126, 127), additionally if probiotic lysate is able to modify the gut microbiome, and finally if both treatments used together or alone can delay the onset of cognitive decline, which is the typical symptom of AD.

The group which received both treatments APP/PS1^{TG}-ExPr showed an improved performance on the MWM test, they outperformed all other transgenic groups and also the WT group (Figure 12.). This test is the most widely used in animals regarding spatial memory and hippocampal function (123). Exercise has been proved to be beneficial for spatial memory preserving in rodents suffering from AD (126, 128). Interestingly only exercise receiving group APP/PS1^{TG}-Ex did not perform well in MWM test. This can be the effect of forced method of exercise, as it was observed previously by the research of Yuede (129). In contrast to the bad performance in spatial memory testing, these animals had the lowest A β plaque number in the hippocampus (as seen in Figure 17.). Low levels of plaques were also described by Chao et al even though they have used exercise in a later stage of disease (130). These results suggest that exercise and probiotic lysate has a diverse effect both on spatial memory and on A β plaque deposition in AD mice (131).

In OFT we did not observe any different area crossing activity among all study groups showing that exercise receiving groups were not more active, they did not move more neither in the inner nor in the outer zones of the arena. This result was also demonstrated with Tg2576 AD mice, exercise do not alter activity level in mice (132).

Also there is no difference in exploratory activity between WT and APP/PS1 groups (Figure 13.A.C.). These findings are in accordance with a study performed in 2018 by Zhang and colleagues, where they investigated the role of environmental enrichment in APP/PS1 mice. They found that enriched environment results in similar exploratory activity in AD mice than in WT mice (133). Interestingly we have found that latency times among the transgenic groups did not differ from each other, but if we compared APP/PS1^{TG}-C to the WT group, we could observe a little sign of anxiety (Figure 13.B.). All transgenic groups had a significantly major latency time than WT group. Similarly, to inner or outer area crossings, we did not find any differences in total exploratory activity.

In NOR test APP/PS1^{TG}-Ex significantly underperformed the APP/PS1^{TG}-C group, control group have spent more time investigating the new object, suggesting the possibility that they have remembered better for the old object than exercise treated group. This result is in contrary to previous and current scientific results, where exercise significantly improved recognition memory in rodents suffering from AD (132, 134). Underlying cause should be closely investigated in a future research. Interestingly APP/PS1^{TG}-ExPr group performed similarly to APP/PS1^{TG}-Pr group indicating that probiotic lysate in this case can overregulate the effects of exercise (Figure 14.).

In spontaneous alternation test WT group outperformed all transgenic groups, they have made almost 100% correct alterations, while transgenic groups were around or less than 75% (Figure 15.A.). This result is in accordance with other described cases (135, 136). If we only observe the transgenic groups no significant differences were shown in their performance (Figure 15.B.). We believe that this test correctly shows that APP/PS1 transgenic mice have serious problems remembering the maze arms where they have previously entered, they have inaccurate navigation skills. These findings are in contrast to previous observations made by the research group of Liu (63). Still combination treatment receiving group outperformed the other transgenic groups.

Both treatments applied alone or in a combination had a serious effect on amyloid plaque quantity and size in the hippocampal arena. Number of amyloid plaques was significantly lower in the APP/PS1^{TG}-Ex group than in APP/PS1^{TG}-C and APP/PS1^{TG}-ExPr group. It seems that exercise alone has a greater effect on amyloid

plaque number. Moreover it was noticed that all treated groups had decreased levels of the area covered by the amyloid plaques in the hippocampal region, suggesting that the probiotic lysate treatment and exercise as well can decrease the plaques size (Figure 17.). In contrary to the lowest number of amyloid plaque depositions, no significant improvement was seen in MWM test in the exercise receiving group. Additionally APP/PS1^{TG}-ExPr group which performed significantly better in the MWM test had the highest number of A β plaques in the hippocampal region. This result suggests that there is no linear correlation between beta-amyloid accumulation and spatial memory impairment in this strain of AD transgenic mice (137).

Microglia investigation resulted in an observation that microglia surrounds the plaques, increased microglia number was counted. This is in agreement with previous work and suggest that microglia accumulation around amyloid plaques may have an important role in A β clearing, however ablation of microglia in APP/PS1 mice did not result in different plaque deposition which raises further questions about the role of microglia in AD pathology (138). We think that this must be the sign for immune reaction because microglia activation is more noticeable around the plaques (21). Unfortunately, we did not measure microglia activation that could have given us some more interesting results.

It was described previously that OGG1 levels are decreased in patients suffering from AD compared to healthy individuals, and this pattern was also present in animal models used to demonstrate AD symptoms (139). Exercise modulates the levels of OGG1 (140) and must upregulate OGG1 (45). Our findings are in accordance with previous studies, OGG1 levels were the highest in the APP/PS1^{TG}-Ex group compared to all transgenic groups, and moreover they were significantly higher than APP/PS1^{TG}-Pr and APP/PS1^{TG}-ExPr groups. Interestingly in the combined group the effects of exercise was not enough to exceed the effects of probiotic lysate.

In the microbiome we have found several interesting results. First of all, we have observed that at genus level Firmicutes and Bacteroidetes genres were most abundantly present in fecal samples. The ratio of Firmicutes/Bacteroidetes was the highest in the WT group while it was the lowest in APP/PS1^{TG}-Pr group (Figure 20.).

On the phylum and species level we observed that exercise increased the number of butyrate producing bacteria (104, 106). Butyrate is produced when bacteria ferments dietary fibers. Butyrate can be helpful to suppress inflammation, and reduced inflammation can result in decreased beta amyloid levels (107), which is exactly what we observed in our study groups. It is interesting that WT mice had significantly higher levels of butyrate producing bacteria compared to APP/PS1^{TG}-C group showing that AD seriously altered the microbiome composition. Investigation between transgenic groups revealed that APP/PS1^{TG}-Ex group had the highest levels of butyrate producing strains such as *Clostridium spp.*, *Eubacterium spp.* and *Roseburia spp.* (Figure 21.C.), compared to other transgenic groups, showing the enormous positive effect of exercise on the GIT. Butyrate is essential for proper mucin production in the intestines. Other epithelial layer modifying bacteria was present in the fecal samples. *Bacterioides spp.* is a known succinate producer which can negatively modify mucin production, also low levels of *Prevotella spp.* suggest problems with mucin production in APP/PS1^{TG}-C group and may have a diagnostic value for gut permeability. WT mice had significantly elevated levels of *Prevotella spp.* in their fecal sample altogether with low levels of *Bacterioides spp.* indicating a better preserved mucin layer (Figure 21.A.). Moreover levels of *Prevotella spp.* has been shown to have an effect on human mood (141), hence AD linked psychological upset could be the result of decreased levels of *Prevotella spp.*

Decreased cognitive function can be in accordance with the elevated presence of *Bacterioides thetaiotaomicron* in transgenic control group compared to WT mice (Figure 22.A.). These bacteria produce acetate and succinate which can be responsible for lowered mucin production. Lowered mucin content can lead to leaky gut syndrome. We observed that *Bacterioides thetaiotaomicron* levels were the lowest in APP/PS1^{TG}-ExPr group which outperformed other transgenic groups in the MWM test, while APP/PS1^{TG}-Pr group had significantly elevated levels of *Bacterioides thetaiotaomicron* and significantly poorer result in MWM test compared to the combined treatment receiving transgenic group (Figure 12. and 22.B.).

We observed that *Lactobacillus johnsonii* levels positively correlated to A β plaque content and distribution. Highest levels were observed in APP/PS1^{TG}-C group (Figure 17. and 22.B.). In addition it was found that *Lactobacillus johnsonii* which has well-documented antimicrobial properties, generates excessive H₂O₂ (142). Exercise is a

well known oxidative stress reducer and our data underlies this result, APP/PS1^{TG}-Ex group had the lowest level of these bacteria (Figure 21.D.).

B12 producing *Lactobacillus reuteri* was also present in our samples. *Lactobacillus reuteri* has the highest levels in probiotic lysate treatment receiving groups APP/PS1^{TG}-Pr and APP/PS1^{TG}-ExPr (Figure 22.B.). This result demonstrates that probiotic lysate is able to modify the gut microbiome in a positive way since B12 vitamin is essential for normal brain function and decreased presence may lead to AD (114).

6. Conclusions

At the objectives chapter I presented the aims of the study, now we need to review the 4 hypotheses I made:

1. Regular physical activity and probiotic lysate treatment will enhance the cognitive function in AD transgenic mice. **PARTLY TRUE** We have seen better performance in the MWM test from APP/PS1^{TG}-ExPr group. On the contrary we did not observe any major changes in other cognitive test compared to the WT group.
2. Our aim was to illustrate that the suggested beneficial effects of exercise and probiotic treatment have different mechanisms, therefore these effects can be summarized and thus reduce the accumulation of Aβ plaques. **PARTLY TRUE** We have measured lower levels of plaques in treated groups, and we observed different mechanisms of how treatments work but these effects in the end could not be summarized, they acted in different ways.
3. Training and/or probiotic lysate treatment will positively modify gut microbiome composition. **PARTLY TRUE** Both treatments modified the gut microbiome, but since we do not know the exact mechanism how bacterial products affect the brain, we cannot say with absolute certainty that modifications were positive.
4. HIIT will have a positive effect on cognition and will cause a delayed progression of AD. **TRUE** We could observe better performance in MWM test, elevated OGG1 levels and reduced plaque area.

7. Summary

7.1. Summary in English

There was an emerging interest lately for the function of the gut-brain axis and for the possible preventive methods for AD. For this reason, we investigated the effects of interval training and probiotic lysate supplementation on transgenic AD mice.

Exercise seems to be a preventive method for AD because it not only preserved the spatial memory of the mice but decreased the amyloid levels in the hippocampus. Moreover, OGG1 levels were elevated in the brain. In the microbiome of exercised animals, we could measure elevated levels of butyrate-producing bacteria, while on the contrary *Prevotella spp* was present only at a lower level. Further investigation is needed to decide which changes have a major effect on brain function. Also, we should investigate that leaky gut syndrome can underlie AD.

In our study we have used a probiotic lysate (heat-killed bacteria) which of course do not mimic exactly the role of living bacteria, still we observed several positive effects on the microbiome. B12 vitamin-producing bacteria were present at an elevated level in APP/PS1^{TG}-Pr group. Levels of *Lactobacillus spp* increased while *Clostridium* levels decreased which can be the effect of Omega 3 fatty acids. Surprisingly probiotic lysate decreased the level of butyrate-producing bacteria. Unfortunately, on cognition, we could not measure significant improvements thanks to probiotic lysate usage.

In summary: both treatments had beneficial effects on the course of AD, mostly in cognition and microbial composition. We could prove the advantages of exercise with better cognition, decreased plaque number, elevated OGG1 level and with an elevated number in butyrate-producing bacteria. Probiotic lysate did not have as many positive effects as exercise, but an elevation in *Lactobacillus spp* could be observed which can serve as a substrate for other beneficial bacteria. Future direction is to find the exact mechanism on the microbial-brain communication and to find all the products secreted by bacteria which can be in connection with AD either positively or negatively hence creating the opportunity to find an effective but non-invasive treatment for patients suffering from this disease.

7.2. Summary in Hungarian- Összefoglalás

Az utóbbi időkben nagy az érdeklődés a bélflóra-agy kapcsolatára valamint az Alzheimer kór lehetséges prevenciók eszközeire vonatkozóan. Éppen ezért vizsgáltuk az interval típusú edzés és egy lizált probiotikumokat tartalmazó táplálék kiegészítő hatását Alzheimer kóros transzgenikus egereken.

Az edzés jó prevenciók eszköznek tűnik, mert azon felül, hogy megőrizte a térbeli memóriát a transzgenikus állatokban megfigyelhettük az amyloid plakkok csökkenő számát a hippocampusban. Ezen felül az agy vizsgált területein emelkedett OGG1 szinttel találkozhattunk. Ha az edzésben részesített állatok mikrobiomját vizsgáljuk, megfigyelhetjük a butirát termelő baktériumok emelkedett szintjét, miközben a Prevotellák száma csökkenő tendenciát mutatott. Azonban további vizsgálatok szükségesek ahhoz, hogy megállapítsuk, mely változásoknak van a legnagyobb hatása az agyi funkciókra. Ezen felül azt is érdemes lenne megvizsgálni, hogy az áteresztő bél szindróma okozhat-e Alzheimer kórt.

Kísérletünkben hővel elölt baktériumokat probiotikum lizátumot használtunk, melynek természetesen nincs ugyanolyan hatása, mintha élő baktériumokat használtunk volna. Ennek ellenére észrevettünk pozitív változásokat a mikrobiomban. B12 termelő baktérium emelkedett mennyiségét találtuk a APP/PS1^{TG}-Pr csoportban. A lactobacillusok száma emelkedett, míg a Clostridiumok száma csökkent ebben a csoportban mely az Omega 3 zsírsavak hatása is lehet. Meglepetésünkre a probiotikum csökkentette a butirát termelő baktériumok számát és sajnos a kognitív funkciókban sem láttunk akkora javulást mint vártuk.

Összességében kijelenthetjük, hogy mindkét típusú kezelésnek voltak előnyei az észlelésre és a mikrobiomra nézve is. Az edzés pozitív hatásai javuló észlelésben, csökkenő amyloid plakk számban emelkedő OGG1 értékben és a butirát termelő baktériumok magas számában mutatkozott meg. A probiotikus lizátumnak nem volt annyi pozitív hatása mint az edzésnek, de a Lactobacillusok emelkedett aránya jó táptalaja lehet egyéb kedvező hatású baktérium számára. A jövőbeni kutatás célja az lenne, hogy feltérképezzük a pontos mechanizmust a bél-agy kommunikációban, és hogy megtaláljuk az összes, baktériumok által kiválasztott terméket, melyek hatással

lehetnek az AD-re akár pozitívan akár negatívan, így képesek lennénk kifejleszteni egy hatásos de nem invazív terápiát a betegségben szenvedők számára.

8. Bibliography

1. Alzheimer A, Stelzmann RA, Schnitzlein HN, Murtagh FR. (1995) An English translation of Alzheimer's 1907 paper, "Über eine eigenartige Erkrankung der Hirnrinde". Clin Anat, 8: 429-431.
2. Alzheimer's A. (2016) 2016 Alzheimer's disease facts and figures. Alzheimers Dement, 12: 459-509.
3. Jack CR, Jr., Albert MS, Knopman DS, McKhann GM, Sperling RA, Carrillo MC, Thies B, Phelps CH. (2011) Introduction to the recommendations from the National Institute on Aging-Alzheimer's Association workgroups on diagnostic guidelines for Alzheimer's disease. Alzheimers Dement, 7: 257-262.
4. Jin WS, Bu XL, Wang YR, Li L, Li WW, Liu YH, Zhu C, Yao XQ, Chen Y, Gao CY, Zhang T, Zhou HD, Zeng F, Wang YJ. (2017) Reduced Cardiovascular Functions in Patients with Alzheimer's Disease. J Alzheimers Dis, 58: 919-925.
5. Lane CA, Hardy J, Schott JM. (2018) Alzheimer's disease. Eur J Neurol, 25: 59-70.
6. Smith MA. (1998) Alzheimer disease. Int Rev Neurobiol, 42: 1-54.
7. Apostolova LG, Green AE, Babakchanian S, Hwang KS, Chou YY, Toga AW, Thompson PM. (2012) Hippocampal atrophy and ventricular enlargement in normal aging, mild cognitive impairment (MCI), and Alzheimer Disease. Alzheimer Dis Assoc Disord, 26: 17-27.
8. American Psychiatric A, American Psychiatric A, Force DSMT. Diagnostic and statistical manual of mental disorders. 2013.
9. McGeer PL, McGeer EG. (2013) The amyloid cascade-inflammatory hypothesis of Alzheimer disease: implications for therapy. Acta Neuropathol, 126: 479-497.
10. Patterson C: 2018. World Alzheimer Report 2018 The state of the art of dementia research:New frontiers. London: Alzheimer's Disease International (ADI),; accessed 2018 September 2018 <https://www.alz.co.uk/research/WorldAlzheimerReport2018.pdf>.
11. Masters CL, Bateman R, Blennow K, Rowe CC, Sperling RA, Cummings JL. (2015) Alzheimer's disease. Nat Rev Dis Primers, 1: 15056.

12. Pike CJ. (2017) Sex and the development of Alzheimer's disease. *J Neurosci Res*, 95: 671-680.
13. Lerch JP, Pruessner JC, Zijdenbos A, Hampel H, Teipel SJ, Evans AC. (2005) Focal decline of cortical thickness in Alzheimer's disease identified by computational neuroanatomy. *Cereb Cortex*, 15: 995-1001.
14. Fonseca Ana Catarina RG, Resende R, Cardoso Sandra M, Pereira Cláudia F: The role of proteotoxic stress in vascular dysfunction in the pathogenesis of Alzheimer's disease. In: *Endoplasmic Reticulum Stress in Diseases*. vol. 2; 2015.
15. Hardy JA, Higgins GA. (1992) Alzheimer's disease: the amyloid cascade hypothesis. *Science*, 256: 184-185.
16. Ricciarelli R, Fedele E. (2017) The Amyloid Cascade Hypothesis in Alzheimer's Disease: It's Time to Change Our Mind. *Curr Neuropharmacol*, 15: 926-935.
17. Holtzman DM, Bales KR, Paul SM, DeMattos RB. (2002) Abeta immunization and anti-Abeta antibodies: potential therapies for the prevention and treatment of Alzheimer's disease. *Adv Drug Deliv Rev*, 54: 1603-1613.
18. Thinakaran G, Koo EH. (2008) Amyloid precursor protein trafficking, processing, and function. *J Biol Chem*, 283: 29615-29619.
19. Wang R, Sweeney D, Gandy SE, Sisodia SS. (1996) The profile of soluble amyloid beta protein in cultured cell media. Detection and quantification of amyloid beta protein and variants by immunoprecipitation-mass spectrometry. *J Biol Chem*, 271: 31894-31902.
20. Chow VW, Mattson MP, Wong PC, Gleichmann M. (2010) An overview of APP processing enzymes and products. *Neuromolecular Med*, 12: 1-12.
21. Gomez-Isla T, Growdon WB, McNamara MJ, Nochlin D, Bird TD, Arango JC, Lopera F, Kosik KS, Lantos PL, Cairns NJ, Hyman BT. (1999) The impact of different presenilin 1 and presenilin 2 mutations on amyloid deposition, neurofibrillary changes and neuronal loss in the familial Alzheimer's disease brain: evidence for other phenotype-modifying factors. *Brain*, 122 (Pt 9): 1709-1719.

22. Sisodia SS, St George-Hyslop PH. (2002) gamma-Secretase, Notch, Abeta and Alzheimer's disease: where do the presenilins fit in? *Nat Rev Neurosci*, 3: 281-290.
23. National Institute of Health: 2013. PSEN1 gene. accessed 20 April, 2019. <https://ghr.nlm.nih.gov/gene/PSEN1>.
24. Marr RA, Hafez DM. (2014) Amyloid-beta and Alzheimer's disease: the role of neprilysin-2 in amyloid-beta clearance. *Front Aging Neurosci*, 6: 187.
25. Turner AJ, Isaac RE, Coates D. (2001) The neprilysin (NEP) family of zinc metalloendopeptidases: genomics and function. *Bioessays*, 23: 261-269.
26. Hafez D, Huang JY, Huynh AM, Valtierra S, Rockenstein E, Bruno AM, Lu B, DesGroseillers L, Masliah E, Marr RA. (2011) Neprilysin-2 is an important beta-amyloid degrading enzyme. *Am J Pathol*, 178: 306-312.
27. Kim J, Basak JM, Holtzman DM. (2009) The role of apolipoprotein E in Alzheimer's disease. *Neuron*, 63: 287-303.
28. Liu S, Park S, Allington G, Prelli F, Sun Y, Marta-Ariza M, Scholtzova H, Biswas G, Brown B, Verghese PB, Mehta PD, Kwon YU, Wisniewski T. (2017) Targeting Apolipoprotein E/Amyloid beta Binding by Peptoid CPO_Abeta17-21 P Ameliorates Alzheimer's Disease Related Pathology and Cognitive Decline. *Sci Rep*, 7: 8009.
29. Zempel H, Mandelkow E. (2014) Lost after translation: missorting of Tau protein and consequences for Alzheimer disease. *Trends Neurosci*, 37: 721-732.
30. Brion JP. (1998) Neurofibrillary tangles and Alzheimer's disease. *Eur Neurol*, 40: 130-140.
31. Mohandas E, Rajmohan V, Raghunath B. (2009) Neurobiology of Alzheimer's disease. *Indian J Psychiatry*, 51: 55-61.
32. Binder LI, Guillozet-Bongaarts AL, Garcia-Sierra F, Berry RW. (2005) Tau, tangles, and Alzheimer's disease. *Biochim Biophys Acta*, 1739: 216-223.
33. Abcam: 2020. Beta-amyloid and tau in Alzheimer's disease. accessed 2020 January 15, 2020. <https://www.abcam.com/neuroscience/beta-amyloid-and-tau-in-alzheimers-disease>.

34. Bryan KJ, Lee H, Perry G, Smith MA, Casadesus G. Transgenic Mouse Models of Alzheimer's Disease: Behavioral Testing and Considerations. In. Edited by Edito, *Methods of Behavior Analysis in Neuroscience* Boca Raton (FL), 2009.
35. Games D, Adams D, Alessandrini R, Barbour R, Berthelette P, Blackwell C, Carr T, Clemens J, Donaldson T, Gillespie F, et al. (1995) Alzheimer-type neuropathology in transgenic mice overexpressing V717F beta-amyloid precursor protein. *Nature*, 373: 523-527.
36. Hsiao K, Chapman P, Nilsen S, Eckman C, Harigaya Y, YOUNKIN S, Yang F, Cole G. (1996) Correlative memory deficits, A β elevation, and amyloid plaques in transgenic mice. *Science*, 274: 99-102.
37. Shen J, Bronson RT, Chen DF, Xia W, Selkoe DJ, Tonegawa S. (1997) Skeletal and CNS defects in Presenilin-1-deficient mice. *Cell*, 89: 629-639.
38. Elder GA, Gama Sosa MA, De Gasperi R. (2010) Transgenic mouse models of Alzheimer's disease. *Mt Sinai J Med*, 77: 69-81.
39. Kitazawa M, Medeiros R, Laferla FM. (2012) Transgenic mouse models of Alzheimer disease: developing a better model as a tool for therapeutic interventions. *Curr Pharm Des*, 18: 1131-1147.
40. Perry G, Cash AD, Smith MA. (2002) Alzheimer Disease and Oxidative Stress. *J Biomed Biotechnol*, 2: 120-123.
41. Pocernich CB, Butterfield DA. (2012) Elevation of glutathione as a therapeutic strategy in Alzheimer disease. *Biochim Biophys Acta*, 1822: 625-630.
42. Chen Z, Zhong C. (2014) Oxidative stress in Alzheimer's disease. *Neurosci Bull*, 30: 271-281.
43. Tonnies E, Trushina E. (2017) Oxidative Stress, Synaptic Dysfunction, and Alzheimer's Disease. *J Alzheimers Dis*, 57: 1105-1121.
44. Ali T, Kim T, Rehman SU, Khan MS, Amin FU, Khan M, Ikram M, Kim MO. (2018) Natural Dietary Supplementation of Anthocyanins via PI3K/Akt/Nrf2/HO-1 Pathways Mitigate Oxidative Stress, Neurodegeneration, and Memory Impairment in a Mouse Model of Alzheimer's Disease. *Mol Neurobiol*, 55: 6076-6093.
45. Boldogh I, Hajas G, Aguilera-Aguirre L, Hegde ML, Radak Z, Bacsı A, Sur S, Hazra TK, Mitra S. (2012) Activation of ras signaling pathway by 8-oxoguanine

- DNA glycosylase bound to its excision product, 8-oxoguanine. *J Biol Chem*, 287: 20769-20773.
46. Gackowski D, Rozalski R, Siomek A, Dziaman T, Nicpon K, Klimarczyk M, Araszkiewicz A, Olinski R. (2008) Oxidative stress and oxidative DNA damage is characteristic for mixed Alzheimer disease/vascular dementia. *J Neurol Sci*, 266: 57-62.
 47. Dincer Y, Akkaya C, Mutlu T, Yavuzer S, Erkol G, Bozluolcay M, Guven M. (2019) DNA repair gene OGG1 polymorphism and its relation with oxidative DNA damage in patients with Alzheimer's disease. *Neurosci Lett*, 709: 134362.
 48. Lee CY, Landreth GE. (2010) The role of microglia in amyloid clearance from the AD brain. *J Neural Transm (Vienna)*, 117: 949-960.
 49. Wegiel J, Wang KC, Tarnawski M, Lach B. (2000) Microglia cells are the driving force in fibrillar plaque formation, whereas astrocytes are a leading factor in plaque degradation. *Acta Neuropathol*, 100: 356-364.
 50. Kim YS, Joh TH. (2006) Microglia, major player in the brain inflammation: their roles in the pathogenesis of Parkinson's disease. *Exp Mol Med*, 38: 333-347.
 51. Hansen DV, Hanson JE, Sheng M. (2018) Microglia in Alzheimer's disease. *J Cell Biol*, 217: 459-472.
 52. van Praag H. (2008) Neurogenesis and exercise: past and future directions. *Neuromolecular Med*, 10: 128-140.
 53. van Praag H, Christie BR, Sejnowski TJ, Gage FH. (1999) Running enhances neurogenesis, learning, and long-term potentiation in mice. *Proc Natl Acad Sci U S A*, 96: 13427-13431.
 54. Hu J, Imam SZ, Hashiguchi K, de Souza-Pinto NC, Bohr VA. (2005) Phosphorylation of human oxoguanine DNA glycosylase (alpha-OGG1) modulates its function. *Nucleic Acids Res*, 33: 3271-3282.
 55. Marton O, Koltai E, Takeda M, Mimura T, Pajk M, Abraham D, Koch LG, Britton SL, Higuchi M, Boldogh I, Radak Z. (2016) The rate of training response to aerobic exercise affects brain function of rats. *Neurochem Int*, 99: 16-23.

56. Scarmeas N, Luchsinger JA, Schupf N, Brickman AM, Cosentino S, Tang MX, Stern Y. (2009) Physical activity, diet, and risk of Alzheimer disease. *JAMA*, 302: 627-637.
57. Ballard C, Khan Z, Clack H, Corbett A. (2011) Nonpharmacological treatment of Alzheimer disease. *Can J Psychiatry*, 56: 589-595.
58. Radak Z, Hart N, Sarga L, Koltai E, Atalay M, Ohno H, Boldogh I. (2010) Exercise plays a preventive role against Alzheimer's disease. *J Alzheimers Dis*, 20: 777-783.
59. Leasure JL, Jones M. (2008) Forced and voluntary exercise differentially affect brain and behavior. *Neuroscience*, 156: 456-465.
60. Wewege M, van den Berg R, Ward RE, Keech A. (2017) The effects of high-intensity interval training vs. moderate-intensity continuous training on body composition in overweight and obese adults: a systematic review and meta-analysis. *Obes Rev*, 18: 635-646.
61. Um HS, Kang EB, Leem YH, Cho IH, Yang CH, Chae KR, Hwang DY, Cho JY. (2008) Exercise training acts as a therapeutic strategy for reduction of the pathogenic phenotypes for Alzheimer's disease in an NSE/APPsw-transgenic model. *Int J Mol Med*, 22: 529-539.
62. Adlard PA, Perreau VM, Pop V, Cotman CW. (2005) Voluntary exercise decreases amyloid load in a transgenic model of Alzheimer's disease. *J Neurosci*, 25: 4217-4221.
63. Liu HL, Zhao G, Cai K, Zhao HH, Shi LD. (2011) Treadmill exercise prevents decline in spatial learning and memory in APP/PS1 transgenic mice through improvement of hippocampal long-term potentiation. *Behav Brain Res*, 218: 308-314.
64. Xu ZQ, Zhang LQ, Wang Q, Marshall C, Xiao N, Gao JY, Wu T, Ding J, Hu G, Xiao M. (2013) Aerobic exercise combined with antioxidative treatment does not counteract moderate- or mid-stage Alzheimer-like pathophysiology of APP/PS1 mice. *CNS Neurosci Ther*, 19: 795-803.
65. Allen JM, Mailing LJ, Niemi GM, Moore R, Cook MD, White BA, Holscher HD, Woods JA. (2018) Exercise Alters Gut Microbiota Composition and Function in Lean and Obese Humans. *Med Sci Sports Exerc*, 50: 747-757.

66. Lazarov O, Robinson J, Tang YP, Hairston IS, Korade-Mirnic Z, Lee VM, Hersh LB, Sapolsky RM, Mirnic K, Sisodia SS. (2005) Environmental enrichment reduces Abeta levels and amyloid deposition in transgenic mice. *Cell*, 120: 701-713.
67. Pang TY, Hannan AJ. (2013) Enhancement of cognitive function in models of brain disease through environmental enrichment and physical activity. *Neuropharmacology*, 64: 515-528.
68. van Praag H, Kempermann G, Gage FH. (2000) Neural consequences of environmental enrichment. *Nat Rev Neurosci*, 1: 191-198.
69. Forsythe P, Kunze WA, Bienenstock J. (2012) On communication between gut microbes and the brain. *Curr Opin Gastroenterol*, 28: 557-562.
70. Gill SR, Pop M, Deboy RT, Eckburg PB, Turnbaugh PJ, Samuel BS, Gordon JI, Relman DA, Fraser-Liggett CM, Nelson KE. (2006) Metagenomic analysis of the human distal gut microbiome. *Science*, 312: 1355-1359.
71. Sender R, Fuchs S, Milo R. (2016) Revised Estimates for the Number of Human and Bacteria Cells in the Body. *PLoS Biol*, 14: e1002533.
72. Hill JM, Clement C, Pogue AI, Bhattacharjee S, Zhao Y, Lukiw WJ. (2014) Pathogenic microbes, the microbiome, and Alzheimer's disease (AD). *Front Aging Neurosci*, 6: 127.
73. Grenham S, Clarke G, Cryan JF, Dinan TG. (2011) Brain-gut-microbe communication in health and disease. *Front Physiol*, 2: 94.
74. Satokari R, Gronroos T, Laitinen K, Salminen S, Isolauri E. (2009) Bifidobacterium and Lactobacillus DNA in the human placenta. *Lett Appl Microbiol*, 48: 8-12.
75. Palmer C, Bik EM, DiGiulio DB, Relman DA, Brown PO. (2007) Development of the human infant intestinal microbiota. *PLoS Biol*, 5: e177.
76. Knol J, Scholtens P, Kafka C, Steenbakkers J, Gro S, Helm K, Klarczyk M, Schopfer H, Bockler HM, Wells J. (2005) Colon microflora in infants fed formula with galacto- and fructo-oligosaccharides: more like breast-fed infants. *J Pediatr Gastroenterol Nutr*, 40: 36-42.
77. Tamburini S, Shen N, Wu HC, Clemente JC. (2016) The microbiome in early life: implications for health outcomes. *Nat Med*, 22: 713-722.

78. Caporaso JG, Lauber CL, Costello EK, Berg-Lyons D, Gonzalez A, Stombaugh J, Knights D, Gajer P, Ravel J, Fierer N, Gordon JI, Knight R. (2011) Moving pictures of the human microbiome. *Genome Biol*, 12: R50.
79. University of Utah: 2019. Your Changing Microbiome. accessed April 27, 2019. <https://learn.genetics.utah.edu/content/microbiome/changing/>.
80. Lloyd-Price J, Abu-Ali G, Huttenhower C. (2016) The healthy human microbiome. *Genome Med*, 8: 51.
81. David LA, Materna AC, Friedman J, Campos-Baptista MI, Blackburn MC, Perrotta A, Erdman SE, Alm EJ. (2014) Host lifestyle affects human microbiota on daily timescales. *Genome Biol*, 15: R89.
82. Dinan TG, Cryan JF. (2017) The Microbiome-Gut-Brain Axis in Health and Disease. *Gastroenterol Clin North Am*, 46: 77-89.
83. Kang DW, Park JG, Ilhan ZE, Wallstrom G, Labaer J, Adams JB, Krajmalnik-Brown R. (2013) Reduced incidence of Prevotella and other fermenters in intestinal microflora of autistic children. *PLoS One*, 8: e68322.
84. Sampson TR, Debelius JW, Thron T, Janssen S, Shastri GG, Ilhan ZE, Challis C, Schretter CE, Rocha S, Gradinaru V, Chesselet MF, Keshavarzian A, Shannon KM, Krajmalnik-Brown R, Wittung-Stafshede P, Knight R, Mazmanian SK. (2016) Gut Microbiota Regulate Motor Deficits and Neuroinflammation in a Model of Parkinson's Disease. *Cell*, 167: 1469-1480 e1412.
85. Cani PD. (2018) Human gut microbiome: hopes, threats and promises. *Gut*, 67: 1716-1725.
86. Sanna S, van Zuydam NR, Mahajan A, Kurilshikov A, Vich Vila A, Vosa U, Mujagic Z, Masclee AAM, Jonkers D, Oosting M, Joosten LAB, Netea MG, Franke L, Zhernakova A, Fu J, Wijmenga C, McCarthy MI. (2019) Causal relationships among the gut microbiome, short-chain fatty acids and metabolic diseases. *Nat Genet*, 51: 600-605.
87. Thomas S, Izard J, Walsh E, Batich K, Chongsathidkiet P, Clarke G, Sela DA, Muller AJ, Mullin JM, Albert K, Gilligan JP, DiGuilio K, Dilbarova R, Alexander W, Prendergast GC. (2017) The Host Microbiome Regulates and

- Maintains Human Health: A Primer and Perspective for Non-Microbiologists. *Cancer Res*, 77: 1783-1812.
88. Maurice CF, Haiser HJ, Turnbaugh PJ. (2013) Xenobiotics shape the physiology and gene expression of the active human gut microbiome. *Cell*, 152: 39-50.
 89. Kashyap PC, Chia N, Nelson H, Segal E, Elinav E. (2017) Microbiome at the Frontier of Personalized Medicine. *Mayo Clin Proc*, 92: 1855-1864.
 90. Camilleri M. (2019) Leaky gut: mechanisms, measurement and clinical implications in humans. *Gut*, 68: 1516-1526.
 91. Ahmad R, Sorrell MF, Batra SK, Dhawan P, Singh AB. (2017) Gut permeability and mucosal inflammation: bad, good or context dependent. *Mucosal Immunol*, 10: 307-317.
 92. Wrzosek L, Miquel S, Noordine ML, Bouet S, Joncquel Chevalier-Curt M, Robert V, Philippe C, Bridonneau C, Cherbuy C, Robbe-Masselot C, Langella P, Thomas M. (2013) *Bacteroides thetaiotaomicron* and *Faecalibacterium prausnitzii* influence the production of mucus glycans and the development of goblet cells in the colonic epithelium of a gnotobiotic model rodent. *BMC Biol*, 11: 61.
 93. Perry RJ, Peng L, Barry NA, Cline GW, Zhang D, Cardone RL, Petersen KF, Kibbey RG, Goodman AL, Shulman GI. (2016) Acetate mediates a microbiome-brain-beta-cell axis to promote metabolic syndrome. *Nature*, 534: 213-217.
 94. Yang BG, Hur KY, Lee MS. (2017) Alterations in Gut Microbiota and Immunity by Dietary Fat. *Yonsei Med J*, 58: 1083-1091.
 95. Obrenovich MEM. (2018) Leaky Gut, Leaky Brain? *Microorganisms*, 6:
 96. Sudo N, Chida Y, Aiba Y, Sonoda J, Oyama N, Yu XN, Kubo C, Koga Y. (2004) Postnatal microbial colonization programs the hypothalamic-pituitary-adrenal system for stress response in mice. *J Physiol*, 558: 263-275.
 97. Saulnier DM, Ringel Y, Heyman MB, Foster JA, Bercik P, Shulman RJ, Versalovic J, Verdu EF, Dinan TG, Hecht G, Guarner F. (2013) The intestinal microbiome, probiotics and prebiotics in neurogastroenterology. *Gut Microbes*, 4: 17-27.

98. Neufeld KM, Kang N, Bienenstock J, Foster JA. (2011) Reduced anxiety-like behavior and central neurochemical change in germ-free mice. *Neurogastroenterol Motil*, 23: 255-264, e119.
99. Luo Y, Zeng B, Zeng L, Du X, Li B, Huo R, Liu L, Wang H, Dong M, Pan J, Zheng P, Zhou C, Wei H, Xie P. (2018) Gut microbiota regulates mouse behaviors through glucocorticoid receptor pathway genes in the hippocampus. *Transl Psychiatry*, 8: 187.
100. Cryan JF, O'Mahony SM. (2011) The microbiome-gut-brain axis: from bowel to behavior. *Neurogastroenterol Motil*, 23: 187-192.
101. Bercik P, Collins SM, Verdu EF. (2012) Microbes and the gut-brain axis. *Neurogastroenterol Motil*, 24: 405-413.
102. Mika A, Fleshner M. (2016) Early-life exercise may promote lasting brain and metabolic health through gut bacterial metabolites. *Immunol Cell Biol*, 94: 151-157.
103. O'Sullivan O, Cronin O, Clarke SF, Murphy EF, Molloy MG, Shanahan F, Cotter PD. (2015) Exercise and the microbiota. *Gut Microbes*, 6: 131-136.
104. Matsumoto M, Inoue R, Tsukahara T, Ushida K, Chiji H, Matsubara N, Hara H. (2008) Voluntary running exercise alters microbiota composition and increases n-butyrate concentration in the rat cecum. *Biosci Biotechnol Biochem*, 72: 572-576.
105. Perrin P, Pierre F, Patry Y, Champ M, Berreur M, Pradal G, Bornet F, Meflah K, Menanteau J. (2001) Only fibres promoting a stable butyrate producing colonic ecosystem decrease the rate of aberrant crypt foci in rats. *Gut*, 48: 53-61.
106. Estaki M, Pither J, Baumeister P, Little JP, Gill SK, Ghosh S, Ahmadi-Vand Z, Marsden KR, Gibson DL. (2016) Cardiorespiratory fitness as a predictor of intestinal microbial diversity and distinct metagenomic functions. *Microbiome*, 4: 42.
107. Zhang L, Wang Y, Xiayu X, Shi C, Chen W, Song N, Fu X, Zhou R, Xu YF, Huang L, Zhu H, Han Y, Qin C. (2017) Altered Gut Microbiota in a Mouse Model of Alzheimer's Disease. *J Alzheimers Dis*, 60: 1241-1257.

108. Dalile B, Van Oudenhove L, Vervliet B, Verbeke K. (2019) The role of short-chain fatty acids in microbiota-gut-brain communication. *Nat Rev Gastroenterol Hepatol*,
109. Alcock J, Maley CC, Aktipis CA. (2014) Is eating behavior manipulated by the gastrointestinal microbiota? Evolutionary pressures and potential mechanisms. *Bioessays*, 36: 940-949.
110. National Institute of Health: 2016. Probiotics: In Depth. accessed April 20, 2019. <https://nccih.nih.gov/health/probiotics/introduction.htm>.
111. LeBlanc JG, Laino JE, del Valle MJ, Vannini V, van Sinderen D, Taranto MP, de Valdez GF, de Giori GS, Sesma F. (2011) B-group vitamin production by lactic acid bacteria--current knowledge and potential applications. *J Appl Microbiol*, 111: 1297-1309.
112. Perkins JB, Pero J. Vitamin Biosynthesis. In. Edited by Editio, *Bacillus subtilis and Its Closest Relatives*. American Society of Microbiology 2002.
113. Taranto MP, Vera JL, Hugenholtz J, De Valdez GF, Sesma F. (2003) *Lactobacillus reuteri* CRL1098 produces cobalamin. *J Bacteriol*, 185: 5643-5647.
114. Troen AM. (2012) Folate and vitamin B12: function and importance in cognitive development. *Nestle Nutr Inst Workshop Ser*, 70: 161-171.
115. Hill JM, Lukiw WJ. (2015) Microbial-generated amyloids and Alzheimer's disease (AD). *Front Aging Neurosci*, 7: 9.
116. Leblhuber F, Steiner K, Schuetz B, Fuchs D, Gostner JM. (2018) Probiotic Supplementation in Patients with Alzheimer's Dementia - An Explorative Intervention Study. *Curr Alzheimer Res*, 15: 1106-1113.
117. Agahi A, Hamidi GA, Daneshvar R, Hamdieh M, Soheili M, Alinaghipour A, Esmaeili Taba SM, Salami M. (2018) Does Severity of Alzheimer's Disease Contribute to Its Responsiveness to Modifying Gut Microbiota? A Double Blind Clinical Trial. *Front Neurol*, 9: 662.
118. Braniste V, Al-Asmakh M, Kowal C, Anuar F, Abbaspour A, Toth M, Korecka A, Bakocevic N, Ng LG, Kundu P, Gulyas B, Halldin C, Hultenby K, Nilsson H, Hebert H, Volpe BT, Diamond B, Pettersson S. (2014) The gut microbiota

- influences blood-brain barrier permeability in mice. *Sci Transl Med*, 6: 263ra158.
119. Prenderville JA, Kennedy PJ, Dinan TG, Cryan JF. (2015) Adding fuel to the fire: the impact of stress on the ageing brain. *Trends Neurosci*, 38: 13-25.
 120. Dinan TG, Cryan JF. (2017) Gut instincts: microbiota as a key regulator of brain development, ageing and neurodegeneration. *J Physiol*, 595: 489-503.
 121. Barnard ND, Bush AI, Ceccarelli A, Cooper J, de Jager CA, Erickson KI, Fraser G, Kesler S, Levin SM, Lucey B, Morris MC, Squitti R. (2014) Dietary and lifestyle guidelines for the prevention of Alzheimer's disease. *Neurobiol Aging*, 35 Suppl 2: S74-78.
 122. Feher J, Pinter E, Kovacs I, Helyes Z, Kemeny A, Markovics A, Plateroti R, Librando A, Cruciani F. (2014) Irritable eye syndrome: neuroimmune mechanisms and benefits of selected nutrients. *Ocul Surf*, 12: 134-145.
 123. Morris R. (1984) Developments of a water-maze procedure for studying spatial learning in the rat. *J Neurosci Methods*, 11: 47-60.
 124. Leger M, Quiedeville A, Bouet V, Haelewyn B, Boulouard M, Schumann-Bard P, Freret T. (2013) Object recognition test in mice. *Nat Protoc*, 8: 2531-2537.
 125. Aziz RK, Bartels D, Best AA, DeJongh M, Disz T, Edwards RA, Formsma K, Gerdes S, Glass EM, Kubal M, Meyer F, Olsen GJ, Olson R, Osterman AL, Overbeek RA, McNeil LK, Paarmann D, Paczian T, Parrello B, Pusch GD, Reich C, Stevens R, Vassieva O, Vonstein V, Wilke A, Zagnitko O. (2008) The RAST Server: rapid annotations using subsystems technology. *BMC Genomics*, 9: 75.
 126. Hoveida R, Alaei H, Oryan S, Parivar K, Reisi P. (2011) Treadmill running improves spatial memory in an animal model of Alzheimer's disease. *Behav Brain Res*, 216: 270-274.
 127. Pareja-Galeano H, Brioché T, Sanchis-Gomar F, Escrivá C, Dromant M, Gómez-Cabrera MC, Vina J. (2012) [Effects of physical exercise on cognitive alterations and oxidative stress in an APP/PSN1 transgenic model of Alzheimer's disease]. *Rev Esp Geriatr Gerontol*, 47: 198-204.
 128. Zhang L, Tang W, Chao FL, Zhou CN, Jiang L, Zhang Y, Liang X, Tang J, Qi YQ, Yang H, He Q, Zhang SS, Zhu L, Peng Y, Tang Y. (2020) Four-month

- treadmill exercise prevents the decline in spatial learning and memory abilities and the loss of spinophilin-immunoreactive puncta in the hippocampus of APP/PS1 transgenic mice. *Neurobiol Dis*, 136: 104723.
129. Yuede CM, Zimmerman SD, Dong H, Kling MJ, Bero AW, Holtzman DM, Timson BF, Csernansky JG. (2009) Effects of voluntary and forced exercise on plaque deposition, hippocampal volume, and behavior in the Tg2576 mouse model of Alzheimer's disease. *Neurobiol Dis*, 35: 426-432.
 130. Chao F, Jiang L, Zhang Y, Zhou C, Zhang L, Tang J, Liang X, Qi Y, Zhu Y, Ma J, Tang Y. (2018) Stereological Investigation of the Effects of Treadmill Running Exercise on the Hippocampal Neurons in Middle-Aged APP/PS1 Transgenic Mice. *J Alzheimers Dis*, 63: 689-703.
 131. Abraham D, Feher J, Scuderi GL, Szabo D, Dobolyi A, Cservenak M, Juhasz J, Ligeti B, Pongor S, Gomez-Cabrera MC, Vina J, Higuchi M, Suzuki K, Boldogh I, Radak Z. (2019) Exercise and probiotics attenuate the development of Alzheimer's disease in transgenic mice: Role of microbiome. *Exp Gerontol*, 115: 122-131.
 132. Thomas R, Zimmerman SD, Yuede KM, Cirrito JR, Tai LM, Timson BF, Yuede CM. (2020) Exercise Training Results in Lower Amyloid Plaque Load and Greater Cognitive Function in an Intensity Dependent Manner in the Tg2576 Mouse Model of Alzheimer's Disease. *Brain Sci*, 10:
 133. Zhang Y, Wang G, Wang L, Zhao J, Huang R, Xiong Q. (2018) The short-term improvements of enriched environment in behaviors and pathological changes of APP/PS1 mice via regulating cytokines. *Hum Vaccin Immunother*, 14: 2003-2011.
 134. Farzi MA, Sadigh-Eteghad S, Ebrahimi K, Talebi M. (2019) Exercise Improves Recognition Memory and Acetylcholinesterase Activity in the Beta Amyloid-Induced Rat Model of Alzheimer's Disease. *Ann Neurosci*, 25: 121-125.
 135. Zhang X, Zhao F, Wang C, Zhang J, Bai Y, Zhou F, Wang Z, Wu M, Yang W, Guo J, Qi J. (2019) AVP(4-8) Improves Cognitive Behaviors and Hippocampal Synaptic Plasticity in the APP/PS1 Mouse Model of Alzheimer's Disease. *Neurosci Bull*,

136. Xu YJ, Mei Y, Qu ZL, Zhang SJ, Zhao W, Fang JS, Wu J, Yang C, Liu SJ, Fang YQ, Wang Q, Zhang YB. (2018) Ligustilide Ameliorates Memory Deficiency in APP/PS1 Transgenic Mice via Restoring Mitochondrial Dysfunction. *Biomed Res Int*, 2018: 4606752.
137. Kamphuis W, Kooijman L, Schetters S, Orre M, Hol EM. (2016) Transcriptional profiling of CD11c-positive microglia accumulating around amyloid plaques in a mouse model for Alzheimer's disease. *Biochim Biophys Acta*, 1862: 1847-1860.
138. Kraft AW, Hu X, Yoon H, Yan P, Xiao Q, Wang Y, Gil SC, Brown J, Wilhelmsson U, Restivo JL, Cirrito JR, Holtzman DM, Kim J, Pekny M, Lee JM. (2013) Attenuating astrocyte activation accelerates plaque pathogenesis in APP/PS1 mice. *FASEB J*, 27: 187-198.
139. Shao C, Xiong S, Li GM, Gu L, Mao G, Markesbery WR, Lovell MA. (2008) Altered 8-oxoguanine glycosylase in mild cognitive impairment and late-stage Alzheimer's disease brain. *Free Radic Biol Med*, 45: 813-819.
140. Sarga L, Hart N, Koch LG, Britton SL, Hajas G, Boldogh I, Ba X, Radak Z. (2013) Aerobic endurance capacity affects spatial memory and SIRT1 is a potent modulator of 8-oxoguanine repair. *Neuroscience*, 252: 326-336.
141. Lin P, Ding B, Feng C, Yin S, Zhang T, Qi X, Lv H, Guo X, Dong K, Zhu Y, Li Q. (2017) Prevotella and Klebsiella proportions in fecal microbial communities are potential characteristic parameters for patients with major depressive disorder. *J Affect Disord*, 207: 300-304.
142. Pridmore RD, Pittet AC, Praplan F, Cavadini C. (2008) Hydrogen peroxide production by *Lactobacillus johnsonii* NCC 533 and its role in anti-Salmonella activity. *FEMS Microbiol Lett*, 283: 210-215.

9. Bibilography of own Publications

In connection with the thesis:

Abraham D, Feher J, Scuderi GL, Szabo D, Dobolyi A, Cservenak M, Juhasz J, Ligeti B, Pongor S, Gomez-Cabrera MC, Vina J, Higuchi M, Suzuki K, Boldogh I, Radak Z. (2019) Exercise and probiotics attenuate the development of Alzheimer's disease in transgenic mice: Role of microbiome. *Exp Gerontol*, 115: 122-131.

IF: 3.376

Marton O, Koltai E, Takeda M, Mimura T, Pajk M, Abraham D, Koch LG, Britton SL, Higuchi M, Boldogh I, Radak Z. (2016) The rate of training response to aerobic exercise affects brain function of rats. *Neurochem Int*, 99: 16-23.

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Not in connection with the thesis:

Perényi H, Szegeczki V, Horváth G, Hinnah B, Tamás A, Radák Z, Ábrahám D, Zákány R, Reglodi D, Juhász T. (2020) Physical Activity Protects the Pathological Alterations of Alzheimer's Disease Kidneys via the Activation of PACAP and BMP Signaling Pathways. *Frontiers in Cellular Neuroscience*, 14:

IF: 3.921 in 2019

Szegeczki V, Horvath G, Perenyi H, Tamas A, Radak Z, Abraham D, Zakany R, Reglodi D, Juhasz T. (2020) Alzheimer's Disease Mouse as a Model of Testis Degeneration. *Int J Mol Sci*, 21:

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